The Role of Sialic Acid in Opsonin-Dependent and Opsonin-Independent Adhesion of Listeria monocytogenes to Murine Peritoneal Macrophages

Srinivas Maganti  
*University of New Hampshire - Main Campus*

Marcia M. Pierce  
*University of New Hampshire - Main Campus*

Alex Hoffmaster  
*University of New Hampshire - Main Campus*

Frank G. Rodgers  
*University of New Hampshire - Main Campus*

Follow this and additional works at: [https://scholars.unh.edu/nhaes](https://scholars.unh.edu/nhaes)

Part of the [Bacteriology Commons](https://scholars.unh.edu/bacteriology), and the [Immunology of Infectious Disease Commons](https://scholars.unh.edu/immunology)

**Recommended Citation**
The Role of Sialic Acid in Opsonin-Dependent and Opsonin-Independent Adhesion of Listeria monocytogenes to Murine Peritoneal Macrophages

This article is available at University of New Hampshire Scholars' Repository: https://scholars.unh.edu/nhaes/257
The Role of Sialic Acid in Opsonin-Dependent and Opsonin-Independent Adhesion of *Listeria monocytogenes* to Murine Peritoneal Macrophages†

SRINIVAS MAGANTI, MARCIA M. PIERCE,‡ ALEX HOFFMASTER,§ AND FRANK G. RODGERS*

Department of Microbiology, University of New Hampshire, Durham, New Hampshire 03824-2617

Received 11 July 1997/Returned for modification 25 September 1997/Accepted 25 November 1997

The adhesion of listeriae to host cells employs mechanisms which are complex and not well understood. *Listeria monocytogenes* is a facultative intracellular pathogen responsible for meningoencephalitis, septicemia, and abortion in susceptible and immunocompromised individuals. Subsequent to colonization and penetration of the gut epithelium, the organism attaches to resident macrophages and replicates intracellularly, thus evading the humoral immune system of the infected host. The focus of these studies was to investigate the attachment of the organism to murine peritoneal macrophages in an opsonin-dependent and opsonin-independent fashion. Assessment of competitive binding experiments by immunofluorescence and enzyme-linked immunosorbent assays showed that adhesion of the organism to macrophages in the presence or absence of opsonins was inhibited (90%) by N-acetyleneuraminic acid (NAcNeu). In addition, the lectin from *Maackia amurensis*, with affinity for NAcNeu-(2,3)galactose, blocked binding of *L. monocytogenes* to host cells. Oxidation of the surface carbohydrates on the organism by using sodium metaperiodate resulted in a dose-dependent reduction (up to 98%) in adherence to macrophages. Monoclonal antibody to complement receptor 3 did not prevent listeriae from binding to mouse macrophages or from replicating within the infected cells whether or not normal mouse serum was present. Based on our results, we propose the involvement of NAcNeu, a member of the sialic acid group, in the attachment of *L. monocytogenes* to permissive murine macrophages.

Adherence is the necessary first step in the infectious process of many intracellular pathogens (17, 28). The molecular mechanisms by which adherence and subsequent phagocytosis of the pathogen occur either through an opsonin-mediated process, involving complement or antibody and their appropriate receptors on the host cell, or through an opsonin-independent process in which bacterial adhesins recognize and attach to specific host cell receptors are not well understood. However, the receptors involved in opsonin-dependent adherence may also be involved in opsonin-independent adherence, as in the case of group B streptococci (GBS), which recognize and bind to complement receptor type 3 (CR3) in the absence of opsonins (2). Direct bacterial attachment between adhesins and host cell receptors has also been reported for *Legionella pneumophila* (18, 26) as well as for *Listeria monocytogenes* (5, 8, 24).

*L. monocytogenes* is the causative agent of food-borne listeriosis, a disease which primarily affects immunocompromised individuals, pregnant women, and neonates. Clinical manifestations range from mild, flu-like symptoms to meningoencephalitis and septic abortion. The organism has long been recognized as a facultative intracellular pathogen capable of infecting and replicating within a wide variety of cells, including fibroblasts, epithelial cells, hepatocytes, and cells from the mononuclear phagocyte system (1, 4, 12, 33). It has been shown that mononuclear phagocytes constitute the major effector cells of immunity in experimental infections (1, 20). *L. monocytogenes* has been shown to induce the deposition of C3b and its cleavage products iC3b and C3d through ester and amide linkages, resulting in the activation of the alternative pathway of human complement (6, 10). Recent studies indicate that CR3 mediates phagocytosis of *L. monocytogenes* in the presence of opsonins by a population of listericidal macrophages (8). However, the same studies demonstrated that nonlistericidal macrophages used CR3 as a minor binding molecule for listeriae (8). Use of a monoclonal antibody (MAb) directed against CR3 (CD11b/CD18) inhibited killing of *L. monocytogenes* in a dose-dependent manner for listericidal macrophages; indeed, when the MAb was used at high doses, these treated cells became permissive hosts (9). In contrast, the use of anti-CR3 antibody to block adherence and phagocytosis by nonlistericidal, permissive macrophages was largely ineffective. This finding appears to indicate two possible mechanisms of adherence and uptake for this pathogen: an opsonin-dependent mechanism through CR3 in which the organism is killed by the host cell, and an opsonin-independent mechanism through some receptor other than CR3 in which the organism parasitizes the host cell (27).

Although phagocytosis of *L. monocytogenes* by macrophages in the presence of opsonins has been investigated (7, 21), the role of opsonin-independent phagocytosis in the initiation of infection is not clear. The aim of this study was to investigate the interaction of *L. monocytogenes* with permissive murine peritoneal macrophages prior to phagocytosis and to partially characterize the bacterial adhesive molecules responsible for binding *L. monocytogenes* to these cells both in the presence and in the absence of opsonic components of serum.
MATERIALS AND METHODS

Bacterial strains and culture conditions. The fully virulent strain, UNHNY99, of *L. monocytogenes* serotype 1/2b was isolated on blood agar from the cerebrospinal fluid of an infant who died of neonatal meningoencephalitis. This isolate was subcultured once only on Trypticase soy agar and stored as stock cultures frozen at −70°C in 1% serum-sorbitol. Organism virulence was periodically assessed by the fertile hen egg method (3), and the 50% lethal dose in this system remained at approximately 21 CFU throughout the study period. Aliquots thawed from −70°C were plated on Trypticase soy agar and incubated overnight. Colonies were harvested, resuspended in Trypticase soy broth, and cultured for 8.5 h in a shaking incubator at 37°C. Organisms were collected by centrifugation at 10,000 × g, washed with serum-free Hanks’ balanced salt solution (HBSS; Sigma, St. Louis, Mo.), and resuspended in HBSS to give 10^8 CFU/ml prior to the inoculation of macrophages.

Collection and cultivation of murine peritoneal macrophages. BALB/c mice of both sexes were used at 3 to 6 months of age. Animals were housed at the University of New Hampshire animal maintenance facility according to Animal Care and Use Committee guidelines (National Institutes of Health-approved protocol 1995). Murine peritoneal macrophages were elicited by intraperitoneal injection of each mouse with 2 ml of 4% thioglycolate broth aged for a minimum of 3 months. Mice were sacrificed by carbon dioxide asphyxiation 2.5 to 3.5 days poststimulation, and peritoneal exudate cells were extracted by peritoneal lavage performed three times with HBSS containing 5% normal mouse serum in HBSS at 37°C. Organisms were subsequently added to macrophages and allowed to adhere for 1 h. Nonadherent bacteria were removed by washing, and the plates were assayed for bacterial adherence by IFA and ELISA.

Role of CR3 in opsonin-independent and opsonin-dependent adherence. The rat B-cell hybridoma line M1/70, expressing the MAb immunoglobulin G2b (IgG2b) anti-CR3, with specificity for mouse and human CR3 epitopes, was used to investigate the role of CR3 in opsonin-independent and opsonin-dependent adherence of *L. monocytogenes* to murine macrophages. This antibody blocks the binding of iC3b-coated targets to CR3 (9). The M1/70 cell line was cultured in RPMI 1640 containing 10% fetal bovine serum, and the supernatant was collected and concentrated in a Centricon-10 (Amicon, Beverly, Mass.). The concentrated MAb was purified by protein G-Sepharose 4 Fast Flow column chromatography (Pharmacia, Piscataway, N.J.) and eluted with 0.2 M glycine at pH 2.0. Fractions were further concentrated after adjusting pH to neutrality, and the given in Table 3. Bacterial attachment to host cells was determined in the presence of the sugars and lectins in a competitive fashion. For the sodium metaperiodate treatment regimen, the carbohydrate-oxidizing agent was applied to bacterial surfaces in acetate buffer at pH 5.0. Bacteria were washed to remove the treatment reagent prior to the addition to macrophages. Otherwise, all reagents were made in HBSS, adjusted to optimum pH, and filtered sterilized through 0.22-μm-pore-size membranes prior to use. The attachment of *L. monocytogenes* to macrophages in the absence of the treatment regimens outlined in Tables 1 to 3 served as a control for all of these studies. Furthermore, the binding of GBS strain COH31 to macrophages was used as an additional control for the studies on carbohydrate oxidation with sodium metaperiodate at pH 5.0 to demonstrate enzyme activity at this pH. Following inoculation, listeriae were allowed to adhere to macrophages for 1 h, after which the cells were washed three times with HBSS to remove nonadherent bacteria, and the number of bound listeriae was assayed by IFA and ELISA.

<table>
<thead>
<tr>
<th>Agent</th>
<th>Activity</th>
<th>Treatment time (min)</th>
<th>Function</th>
</tr>
</thead>
<tbody>
<tr>
<td>NAc-neuraminic aldolase</td>
<td>10 U/ml</td>
<td>60</td>
<td>Degradation of protein-, sugar-, and lipid-containing moieties on the surface of <em>L. monocytogenes</em></td>
</tr>
<tr>
<td>β-Galactosidase</td>
<td>100 U/ml</td>
<td>60</td>
<td></td>
</tr>
<tr>
<td>Chymotrypsin</td>
<td>500 U/ml</td>
<td>60</td>
<td></td>
</tr>
<tr>
<td>Lipase</td>
<td>100 U/ml</td>
<td>60</td>
<td></td>
</tr>
<tr>
<td>Neuraminidase</td>
<td>20 U/ml</td>
<td>60</td>
<td></td>
</tr>
<tr>
<td>Protease</td>
<td>10 U/ml</td>
<td>60</td>
<td></td>
</tr>
<tr>
<td>Trypsin</td>
<td>250 U/ml</td>
<td>60</td>
<td></td>
</tr>
<tr>
<td>Glutaraldehyde</td>
<td>0.1%</td>
<td>10</td>
<td>Immobilization of protein moieties</td>
</tr>
<tr>
<td>Sodium metaperiodate</td>
<td>500 mM</td>
<td>60</td>
<td>Oxidation of carbohydrate moieties</td>
</tr>
</tbody>
</table>

* Prepared to pH 7.2 in HBSS (except for sodium metaperiodate, which was used at pH 5.0 in acetate buffer, and neuraminidase, used at pH 5.0 in HBSS) and added to bacterial cells for the specified time then washed to remove excess.
protein content was determined by the bicinchoninic acid assay (Pierce, Rockford, Ill.).

Macrophage monolayers were incubated for 1 h with RPMI 1640 medium containing cytochalasin D (0.5 μg/ml) and rat IgG (10 μg/ml) to prevent organism uptake through phagocytosis and to block Fc receptor-mediated binding. Following cytochalasin treatment, macrophage monolayers were incubated at 37°C for 1 h with NAcNeu or for 30 min with increasing concentrations of the purified anti-CR3 MAb ranging from 0.1 to 25 μg/ml prior to the addition of listeria.

Adherence assays. (i) Indirect IFA. Macrophages, washed to remove nonadherent bacteria, were fixed in 10% (vol/vol) formalin in HBSS (pH 7.2) for 30 min at 25°C. After fixing, the cells were wash with phosphate-buffered saline (PBS) and treated with rabbit polyclonal anti-L. monocytogenes antiseraum for 1 h at 37°C. Unbound globulin was removed, and goat anti-rabbit fluorescein iso-thiocyanate-conjugated antibody (Boehringer Mannheim, Indianapolis, Ind.) was added for 1 h at 37°C. Cells were stained with 0.01% propidium iodide in PBS for 20 min, washed, air dried, and mounted in glycerol containing 1% 1,4-diazobicyclo (2,2,2) octane. For the purposes of this study, a 50% decrease compared with untreated controls in the number of bacteria visualized as adherent to macrophages after a particular organism or host cell treatment was considered indicative of an interruption in adhesion of the organism to host cells (16).

(ii) ELISA. Macrophages, washed to remove nonadherent bacteria, were fixed in absolute methanol for 10 min at 25°C and dried at 37°C. The plates were then washed three times with PBS containing 0.05% Tween 20 at pH 7.2, followed by incubation with 1% gelatin in PBS for 1 h at 37°C. After multiple washing with PBS-Tween 20, cells were incubated with rabbit polyclonal anti-L. monocytogenes antibody and probed with goat anti-rabbit horseradish peroxidase-conjugated antibody (Boehringer Mannheim, Indianapolis, Ind.) each for 1 h at 37°C. After washing the cells, the chromogen 3,3,5,5-tetramethylbenzidine and hydrogen peroxide were added. Development was halted at 10 min by addition of 2 M H2SO4, and the plates were read at 405 nm spectrophotometrically on a model MA310 EIA reader (Whittaker M.A. Bioproducts, Walkersville, Md.). ELISA results enumerating bacteria were derived from a previously established standard curve. Furthermore, a similar 50% reduction in adherent bacteria was used to determine the activity of treatment regimens.

Statistical analysis. IFA data were obtained from the average of three trials, counting the number of adherent bacteria on the first 100 cells counted per trial. Each trial consisted of a total of nine assays in which each treatment was formed three times to give a total of nine separate assays. ELISA data were read at 405 nm spectrophotometrically on a model MA310 EIA reader. ELISA data were generated in a similar fashion. Data from each assay method following each treatment were averaged and analyzed by the unpaired Student’s t test, using Abacus StatView software.

RESULTS

Host cell receptor saturation assays. L. monocytogenes bound to murine peritoneal macrophages in the absence of either complement or specific antibody in a wash-resistant manner. Data from these assays indicated that at an MOI of approximately 1,000, host cell receptors were fully saturated with virulent L. monocytogenes. When an MOI of 100 was used, IFA data showed that macrophage host cell receptors bound approximately 19% of the bacterial inoculum whereas nonspecific binding was minimal. These host cell receptor-bacterial adhesin interaction studies showed that an inoculum of 100 organisms per macrophage was optimal for our purposes, and this was used for all subsequent opsonin-independent and opsonin-modified investigations.

Bacterial surface treatment. With the exception of glutaraldehyde, none of the bacterial cell surface-modifying agents listed in Table 1 were shown to decrease the viability of the bacterial population as determined by viable bacterial cell colony counts. However, modification of adhesin molecules on the bacterial surfaces showed that treatment with the carbohydrate-oxidizing agent sodium metaperiodate abolished binding of the organism to macrophages (Fig. 1). For L. monocytogenes, this destruction of binding by sodium metaperiodate was seen to be concentration dependent (Fig. 2). When GBS were treated with various concentrations of sodium metaperiodate, a similar dose-dependent decrease in binding of the organisms to macrophages was observed (Fig. 3). However, unlike Listeria attachment, which was unaffected by pH treatment alone, GBS binding was decreased by 45%. These data demonstrated that while sodium metaperiodate abolished binding, the use of pH 5.0 in the treatment buffers in the absence of the oxidizing agent had no effect on the attachment of listeriae to host cells. In addition, a 40% reduction in listeria binding (60% adhesion) to macrophages was effected following treatment with neuraminidase. With the exception of glutaraldehyde treatment, which showed an increase in binding of greater than 60% compared with control cells, no treatment

### Table 3. Lectins used in competitive inhibition binding studies

<table>
<thead>
<tr>
<th>Lectin source</th>
<th>Common name (lectin)</th>
<th>Oligosaccharide recognition</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Canavalia ensiformis</td>
<td>Jack bean (concanavalin A)</td>
<td>Branched N-linked hexasaccharides</td>
<td>14</td>
</tr>
<tr>
<td>Limulus polyphemus</td>
<td>Horseshoe crab (L. polyphemus agglutinin)</td>
<td>NAcNeu, NeuGNAc</td>
<td>25</td>
</tr>
<tr>
<td>Maackia amurensis</td>
<td>Maackia (M. amurensis agglutinin)</td>
<td>NAcNeu-α(2,3)Gal, NeuGNAc</td>
<td>32</td>
</tr>
<tr>
<td>Triticum vulgare</td>
<td>Wheat germ (wheat germ agglutinin)</td>
<td>Manβ(1,4)GlcNAc-β(1,4)GlcNAc</td>
<td>34</td>
</tr>
</tbody>
</table>

* NAcNeu, N-acetylneuraminic acid; NeuGNAc, N-acetylglucosamine; Gal, galactose; Man, mannose; GlcNAc, N-acetylgalactosamine. Agents were prepared to pH 7.2 in HBSS and added to macrophage host cells for 1 h. Cells were washed to remove excess, and assays were performed in a competitive fashion.
protocol influenced adherence of listeriae to macrophages. These data indicated that the presence of neuraminidase-sensitive carbohydrate residues on the organism surface were involved in wash-resistant attachment to the receptors on the host cell and that bacterial viability was not essential for this binding to occur.

Host cell surface modification. In similar fashion, host cell surface-modifying agents, with the exception of the aldehyde fixatives and Nonidet P-40, did not adversely affect macrophage viability as assayed by cell counts and trypan blue vital staining. Macrophages exposed to those enzymes or oxidizing agents listed in Table 2 showed no significant change in ability to bind the pathogen in an opsonin-independent manner. However, treatment with either glutaraldehyde or formalde-
binding of the organism when the anti-CR3 MAb was used in a blocking fashion but in the absence of organism uptake was seen to be independent of the concentration of MAb used.

**DISCUSSION**

Adherence to host cell surfaces is a necessary first step in infection by intracellular bacterial pathogens such as *L. monocytogenes*. However, it has been shown that this initial step can take place by two different mechanisms: an opsonin-dependent process, in which antibody and/or complement proteins become involved in the complex interaction between bacteria and host cell; and an opsonin-independent process, in which adhesins present on the bacterial cell surface recognize host cell receptors. It is possible that these two processes result in different reactions by the host cell to the bacterium. For *L. monocytogenes*, uptake by listericidal macrophages via the CR3 receptor resulted in death of the organism, whereas uptake by nonlistericidal macrophages occurred through some other recognition factor and resulted in intracellular replication of the organism (8). Adherence of *L. monocytogenes* to host cell membrane-associated structures is therefore a complex interaction.

Following preliminary studies which established the ability of *L. monocytogenes* to bind to macrophages in a wash-resistant manner (24), the rate at which host cell receptors were saturated with listeriae was investigated. In all cases, an MOI of 1,000 resulted in high levels of organism attachment to the receptors on macrophage surface membranes; however, at this inoculum, the level of nonspecific binding of *L. monocytogenes* to culture plates as seen by IFA proved problematic. An MOI of 100 resulted in a reduced background binding while affording the advantage of greater significance to a 50% reduction in binding as a result of a particular treatment. In essence, this resulted in an inhibition of adherence which was defined as the concentration and nature of a treatment agent that reduced the binding of the organism to the host cells by 50% or greater. In this study, the presence of opsonin-independent attachment of *L. monocytogenes* to murine peritoneal macrophages has been established.

Receptor modification studies showed decreased adherence of *L. monocytogenes* following treatment of the host cell surface structures with formaldehyde and glutaraldehyde. Enzyme treatment of the host cell membrane had little or no effect on the adhesion of the organism to macrophages, while Nonidet P-40 and sodium metaperiodate were also ineffective at influencing the microbe-host cell binding phenomenon. These results indicated that the macrophage receptors involved in the attachment of the organism are protein in nature but that these are either sterically protected from the effects of proteolytic enzymes or present in sufficient quantities on the host cell membranes to allow binding of listeriae to occur at numbers similar to that found in the control macrophage cells.

Attachment of bacteria to mammalian host cells is often mediated by sugar-lectin interactions (5, 17, 23). Mannose (27) and polysialic acid (29) have been shown to facilitate the attachment of *Mycobacterium tuberculosis* and *Neisseria meningitidis*, respectively, to host cells, while *Salmonella typhimurium* adheres to intestinally derived cells via the glycoconjugate receptor, Galβ(1-3)GalNAc (15). In the present study, the results of the competitive binding assays using sugars suggested a role for NAcNeu in the binding of *L. monocytogenes* to macrophages. Indeed, a strong inhibition of attachment (90%) was seen in the presence of 100 mM NAcNeu, while other sugars tested had no effect on the adherence process. Data similar to those reported here showed that the addition of NAcNeu in competitive binding assays impaired the adhesion of strains of *Shigella dysenteriae* and *S. flexneri* to epithelial cells (17). Results from the use of lectins in similar binding assays strongly support the involvement of NAcNeu in adhesion. The lectin from *M. amurensis* showed inhibitory effects similar to those of NAcNeu on binding of *L. monocytogenes*; this lectin has a binding affinity for oligosaccharides which possess the terminal NAcNeu-α(2,3)Gal linkage. Wheat germ agglutinin reduced binding of *L. monocytogenes* by almost 50%. This lectin has binding specificity for both NAcNeu and NAcGlu. *L. polyphemus* lectin is known to bind to NAcNeu as well as N-acetylglycolylneuraminic acid. However, this lectin is extremely large, with an aggregate molecular mass of between 35 and 50 kDa, with four major protein bands seen on sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) (25), whereas the lectin from *M. amurensis* is a single polypeptide of 8 to 9 kDa on SDS-PAGE (32). Steric hindrance may therefore be an important factor in preventing the

---

**FIG. 5.** Binding of opsonized *L. monocytogenes* to macrophages following treatment with NAcNeu. Binding data were generated by IFA; duplicate assays evaluated by ELISA gave similar results. Controls were untreated *L. monocytogenes*. Bars represent standard deviation of the mean. Statistical significance: opsonized organisms treated with NAcNeu, *P* < 0.0001. NMS, normal mouse serum.

**FIG. 6.** Binding of nonopsonized (A) and opsonized (B) *L. monocytogenes* to macrophages following blocking of CR3 receptors on the host cells by using MAb M1/70. Controls in panel A consisted of nonopsonized *L. monocytogenes*; controls in panel B consisted of opsonized *L. monocytogenes*. Binding data were generated by ELISA. Rat IgG was used to saturate Fc receptors on the surface of murine macrophages prior to the addition of antibody M1/70. Bars represent standard deviation of the mean. Statistical significance: (A) *P* > 0.2; (B) *P* > 0.5. NMS, normal mouse serum.
competition of L. polyphemus lectin with the adhesins of this organism.

Bacterial surface treatments indicated a 40% decrease in the binding of the organism following treatment with neuraminidase when used at pH 5.0, the optimal pH for the functioning of this enzyme. Oxidation of the bacterial surface with sodium metaperiodate resulted in a dose-dependent reduction in the binding of L. monocytogenes to mouse macrophages. Following treatment with 10, 100, and 500 mM sodium metaperiodate, this microbial adhesion was reduced by 30, 70, and 98%, respectively. Treatment with other enzymes and fixatives did not inhibit adherence of the organism significantly; in contrast, glutaraldehyde appeared to increase binding by 60% as measured by IF. A. This may be due to cross-linking of surface proteins between individual organisms and subsequent binding of these linked listeriae to the cell surface.

Recent studies have indicated that CR3 is involved in the phagocytosis of L. monocytogenes by listericidal macrophages; however, CR3 is not involved in the phagocytosis of the organism by permissive, nonlistericidal macrophages (8, 9). Accordingly, we wished to determine the effect of the anti-CR3 MAb on adherence of L. monocytogenes as measured by our assay. The presence of this MAb had little or no inhibitory effect on the binding of L. monocytogenes to macrophages in the presence or absence of normal mouse serum. In the present study, a reduction of 30% in the binding of L. monocytogenes to nonlisteridal, thioglycolate-elicited macrophages was effected under non-opsonin-mediated conditions using anti-CR3 antibody. These results were similar to those reported by others (7, 8). However, in contrast to their reported results, the presence or absence of normal mouse serum in our system had no effect on binding in that approximately the same number of listeriae bound to both groups of control macrophage cells. The results reported here indicate that adherence and uptake in permissive macrophages occur by mechanisms other than through CR3 mediation. Significantly, NAcNeu at a concentration of 100 mM was highly inhibitory to the binding of opsonized-L. monocytogenes.

In contrast to GBS, L. monocytogenes does not utilize CR3 as a means for adhering to nonbactericidal, permissive macrophages (2, 8). However, the adhesin molecule used by listeriae to initiate the cellular infectious process was identical whether permissive or listericidal macrophages were used as host cells. Attachment in both the presence and absence of opsonins is strongly inhibited by NAcNeu. In addition, treatment of the bacterial cell surface with the carbohydrate-oxidizing agent sodium metaperiodate also inhibited listerial adhesion to macrophages. Based on our results, we propose the involvement of NAcNeu, a member of the sialic acid group, in the attachment of L. monocytogenes to nonlisterical, permissive host cells. The process of attachment occurred through receptors other than CR3 and allowed the intracellular replication of the organism within host cells to proceed.

Entry of facultative intracellular pathogens into cells that are not professionally phagocytic has been described as a multifactorial process and has been studied extensively in Yersinia, Salmonella, and Shigella as well as Listeria. Indeed, it has been shown that entry and subsequent internalization is effected either through a trigger mechanism as is the case with Salmonella and Shigella or via a zipper mechanism as reported for Yersinia and Listeria (11, 19, 22, 30). Efforts to elucidate the nature of the bacterial ligand (invasin protein) as well as the host cell receptor (β1 integrin receptor) involved in the binding and entry of some facultative intracellular bacterial pathogens into cells has met with some success. Indeed, the critical role of the internalin molecule mediating bacterial entry and invasion of epithelial host cells has been demonstrated (13). In addition, E-cadherin, a calcium-dependent cell-to-cell adhesion glycoprotein located on the surface of epithelial cells, was shown to be the receptor for internalin (22). Further studies have shown that the bacterial surface protein p60 may be involved in uptake of L. monocytogenes by transformed nonproliferating epithelial cells, while the internalin proteins InlA and InlB mediate the invasion of these host cells when in an actively multiplying state (31). From the data presented here, it is possible that internalin proteins, in addition to regulating invasion, are involved in facilitating attachment to permissive peritoneal macrophages. However, the nature of the receptors located on macrophage membranes involved in opsonin-independent binding remains unclear. Additional studies are in progress to determine whether bacterial molecules other than internalin are involved in the attachment of L. monocytogenes to permissive macrophages. Understanding the mechanisms by which L. monocytogenes binds to host cells both in the presence and in the absence of opsonins is critical to elucidating the nature of the intracellular parasitism of this organism and may prove to be an important factor in the development of preventative treatment strategies for listeriosis.

ACKNOWLEDGMENTS

This work was supported by U.S. Department of Agriculture Hatch grant NH 333 and by a Central University Research Fund grant from the Office of Sponsored Research, UNH.

We thank Ronald Gibbons (now deceased) for helpful discussion during the course of this work; Jeffrey W. Hixon, Patricia A. Daggett, Kevin P. Richard, and Robert E. Gibson for technical assistance; and Thomas G. Pistole and Mary Smith for hybridoma cell line M1/70 and samples of purified antibody.

REFERENCES


14. Finlay, B. B., and P. Cossart. 1996. Bacterial surface proteins between individual organisms and subsequent binding of these linked listeriae to the cell surface.

15. Finlay, B. B., and P. Cossart. 1996. Bacterial surface proteins between individual organisms and subsequent binding of these linked listeriae to the cell surface.

16. Finlay, B. B., and P. Cossart. 1996. Bacterial surface proteins between individual organisms and subsequent binding of these linked listeriae to the cell surface.

17. Finlay, B. B., and P. Cossart. 1996. Bacterial surface proteins between individual organisms and subsequent binding of these linked listeriae to the cell surface.
of *Listeria monocytogenes* into cells is mediated by internalin, a repeat protein reminiscent of surface antigens from gram-positive cocci. Cell 65:1127–1141.


32. Wang, W. C., and R. D. Cummings. 1988. The immobilized leukagglutinin from the seeds of *Maackia amurensis* bind with high affinity to complex-type Asn-linked oligosaccharides containing terminal sialic acid-linked alpha-2,3 to penultimate galactose residues. J. Biol. Chem. 263:4576–4585.
