

University of New Hampshire

## University of New Hampshire Scholars' Repository

---

Molecular, Cellular and Biomedical Sciences  
Scholarship

Molecular, Cellular and Biomedical Sciences

---

6-1-2014

### Calcium Homeostasis in Escherichia coli: Characterization of Mutants and Genome Expression of MG1655

Muhammad Arif

Josh Howard

Louis S. Tisa

*University of New Hampshire, Durham, louis.tisa@unh.edu*

Follow this and additional works at: [https://scholars.unh.edu/mcbs\\_facpub](https://scholars.unh.edu/mcbs_facpub)

---

#### Recommended Citation

Arif, M. J. Howard, and L.S. Tisa. 2014. Calcium Homeostasis in Escherichia coli: Isolation and characterization of calcium-sensitive mutants and Global Genome Expression of strain MG1655. Bangladesh Journal of Microbiology 31:1-8

This Article is brought to you for free and open access by the Molecular, Cellular and Biomedical Sciences at University of New Hampshire Scholars' Repository. It has been accepted for inclusion in Molecular, Cellular and Biomedical Sciences Scholarship by an authorized administrator of University of New Hampshire Scholars' Repository. For more information, please contact [Scholarly.Communication@unh.edu](mailto:Scholarly.Communication@unh.edu).

## Original Article

# Calcium Homeostasis in *Escherichia coli*: Characterization of Mutants and Genome Expression of MG1655

Muhammad Arif, Josh Howard and Louis S Tisa\*

Department of Molecular, Cellular and Biomedical Sciences, University of New Hampshire, Durham, NH, USA

While the role of calcium ions as secondary messengers has been well described in eukaryotic cells, little is known about calcium homeostasis in bacteria at the physiological and molecular levels. Genetic and genomic approaches were used to address calcium regulation and to identify genes (*cal*) involved in calcium homeostasis. Transposon mutagenesis of *Escherichia coli* generated several calcium-sensitive mutants that fell into three categories: (i) Ca<sup>2+</sup>-sensitive chemotaxis mutants, (ii) Ca<sup>2+</sup>-sensitive cell division mutants, and (iii) Ca<sup>2+</sup>-sensitive mutants that showed no defects in cell division or chemotaxis. The physiological properties of these Ca<sup>2+</sup>-sensitive mutants were determined. Besides calcium-sensitivity to 75 mM calcium, all of the mutants exhibited increased sensitivities to several divalent cations including Ni<sup>2+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Co<sup>2+</sup>, Zn<sup>2+</sup>, Cu<sup>2+</sup>, and Cd<sup>2+</sup>. To identify the *cal* gene sequence in the Ca<sup>2+</sup>-sensitive mutants, the region of the genes fused to the reporter gene (*phoA*) on the transposon Tn*phoA* was amplified by PCR and sequenced. The sites of the gene fusion for three *cal* mutants were at the *fdoG*, *gpt* and *pqi5* genes. The pleiotropic nature for the *cal* mutations suggested that many genes may be globally regulated by calcium. We then investigated global gene expression patterns of wild-type *E. coli* under calcium-depleted (addition of 10 mM EGTA) and calcium-elevated (addition of 75 mM Ca<sup>2+</sup>) conditions as compared to cultures grown under unstressed conditions. A comprehensive transcriptome analysis using macroarrays exhibited a global regulation of diverse genes within the *E. coli* genome during calcium homeostasis.

**Keywords:** Calcium, Homeostasis, *Escherichia coli*, Transposon, Cations, Genome expression profile

## Introduction

*Escherichia coli* tightly regulate cytosolic free calcium at approximately 100 nM, a level similar to that of eukaryotic cells<sup>1</sup>. Calcium ions serve as secondary chemical messengers in eukaryotic cells and tissues including muscle, neural, and cardiovascular tissue. However, the specific role of calcium ions in prokaryotic cells is not as well defined.

The role of calcium ion participation in bacterial behaviour and signal transduction has been established by several lines of evidence<sup>2-7</sup>. Previous studies demonstrate the following observations: (1) Changing cytoplasmic free Ca<sup>2+</sup> levels in cells of *E. coli* alters their behavior<sup>2</sup>. Tumbling ensues following the release of free Ca<sup>2+</sup> that is produced from caged Ca<sup>2+</sup> compounds such as nitr-5 upon irradiation. (2) Changes in bacterial behaviour are correlated with changes in cytoplasmic free Ca<sup>2+</sup> level<sup>4</sup>. Fluorescent Ca<sup>2+</sup> indicator dye fura-2 was used to show that repellents caused a temporary rise in cytoplasmic free Ca<sup>2+</sup> levels, while attractants caused a temporary fall, and the receptor proteins were required for these effects. (3) Some mutants defective in calcium transport have elevated levels of cytoplasmic free Ca<sup>2+</sup>, tumble continuously, and are defective in chemotaxis<sup>3</sup>. (4) Calcium ion channel blockers inhibit *E. coli* chemotaxis<sup>5-6</sup>.

Ca<sup>2+</sup>-stimulated kinase activities have been found in *E. coli*<sup>8-10</sup> and other bacteria<sup>11-12</sup>. Ca<sup>2+</sup>-binding proteins and calmodulin-like proteins have been reported to be present in a wide variety of bacteria<sup>13</sup>. Immunological evidence indicates the presence of three calmodulin-like proteins in *E. coli*, which were suggested to be involved in calcium ion regulation<sup>14</sup>.

The mechanism of calcium entry is unknown. Four genes (*calA*, *calC*, *calD*, and *chaA*) have been identified in *E. coli* that are involved in calcium homeostasis. All three of these loci, A, C, and D, are associated with Ca<sup>2+</sup>/PO<sub>4</sub><sup>2-</sup> symporter activity<sup>15</sup>. The *chaA* gene is involved in Ca<sup>2+</sup>/H<sup>+</sup> antiporter activity<sup>16</sup>. Recently, ATP has been proposed to regulate calcium efflux in *E. coli* through an ATPase<sup>17</sup>.

To explore the regulatory role of calcium, we decided to generate calcium homeostasis mutant strains and physiologically characterize the mutant strains. In addition, our efforts included a comprehensive assessment of the genome-wide regulation of calcium homeostasis in *E. coli* MG1655 wild-type strain to identify candidate calcium homeostasis genes.

## Materials and Methods

Strains, plasmids and phages

All strains of *E. coli* used in this study are derivatives of K12 and are listed in Table 1.

\*Corresponding author:

Louis S Tisa, Department of Molecular, Cellular and Biomedical Sciences, University of New Hampshire, Durham, NH, USA.

E-mail: louis.tisa@unh.edu

**Table 1.** List of bacterial strains

Strain	Phenotype	Reference or Source
CC118	araD139 $\Delta$ (ara,leu)7697 $\Delta$ lacX74 phoA $\Delta$ 20 galE $\square$ galKthirpsErpoBargEamrecAl	Colin and Beckwith, 1985 <sup>18</sup>
Cal102	CC118::TnphoA	This study
Cal117	CC118::TnphoA	This study
Cal134	CC118::TnphoA	This study
Cal512	CC118::TnphoA	This study
Cal504	CC118::TnphoA	This study
Cal719	CC118::TnphoA	This study
Cal526	CC118::TnphoA	This study
Pho43	CC118::TnphoA	This study
PhoA4	CC118::TnphoA	This study
PhoC23	CC118::TnphoA	This study
MG1655	Wildtype K12 strain, sequenced first.	Blattner et al., 1997 <sup>19</sup>
$\ddot{e}$ TnphoA	Tn5 IS50L::phoA (KmR)	Gutierrez et al., 1987 <sup>20</sup>

### Growth conditions

*E. coli* K12 strain CC118 ( $\Delta$ phoA, spectinomycin resistant) was the parental strain that was used for mutagenesis and subsequent generation of calcium-sensitive mutants. Cells were grown and maintained in LB medium (1.0% bacto-peptone, 0.5% NaCl, 0.5% yeast extract) containing the appropriate antibiotics. The calcium-sensitive mutants were designated: Cal117, Cal512, Pho43, Cal102, Cal134, PhoA4, PhoC23, Cal504, Cal719, and Cal526. For cation sensitivity assays, cells were grown in E medium (0.5% bacto-peptone with 120 mM Tris, 70 mM NaCl, 20 mM KCl, 20 mM NH<sub>4</sub>Cl, 3 mM Na<sub>2</sub>SO<sub>4</sub>, 1 mM MgCl<sub>2</sub>, 2 mM ZnCl<sub>2</sub> and 0.4% glycerol) at a pH of 7.8<sup>21</sup>. For motility and chemotaxis assays, cells were grown in tryptone broth consisting of 1% Bacto-tryptone and 0.5% NaCl and incubated at 35°C. In some cases, cells were grown in Vogel-Bonner medium<sup>22</sup> containing the required amino acids at 1 mM and 50 mM glycerol (minimal glycerol medium) or 50 mM DL-lactate (minimal lactate medium).

### Transposon TnphoA and mutagenesis of CC118

The transposon TnphoA was introduced into strain CC118, the parental wild type *E. coli* strain, by phage  $\ddot{e}$  infection ( $\ddot{e}$  TnphoA) according to the method of Manoil and Beckwith<sup>18</sup> at a multiplicity of infection (MOI) of 0.05. Cells of strain CC118 were incubated at 37°C in LB medium containing 10 mM MgSO<sub>4</sub> and 0.4% maltose. Overnight grown cells were harvested by centrifugation and resuspended in 10 mM MgSO<sub>4</sub>. The washed cells were infected with  $\ddot{e}$  TnphoA lysate as described in the next section.

### Selection of calcium-sensitive mutants

For one series, each 100  $\mu$ l sample of cells was infected with 10, 100, 200, or 500  $\mu$ l of lysate. The mixture was incubated at 30°C for 20 min. Following the addition of 1 ml of LB medium, the cells were allowed to outgrow for 30 min at 30°C. The cells were harvested by centrifugation at 6,000 x g for 10 min and

resuspended in E medium containing glycerol (glycerol-E medium). This procedure was repeated twice and the cells were finally resuspended in 1 ml of glycerol-E medium containing 50 mM CaCl<sub>2</sub> and 60  $\mu$ g/ml kanamycin. Following an incubation at 35°C for 1 h, penicillin G (15,000 U) was added and the cells were allowed to incubate for 3 h. Penicillin G was removed by centrifugation at 6,000 x g for 10 min and resuspending the cell pellet in glycerol-E medium. This washing step was repeated twice. The cells were finally resuspended in 1 ml of glycerol-E medium and 100- $\mu$ l samples were plated on glycerol-E medium plates containing 5 mM CaCl<sub>2</sub>, 60  $\mu$ g/ml kanamycin and 40  $\mu$ g/ml 5-bromo-4-chloro-3-indolyl phosphate (XP) to screen for TnphoA insertions exhibiting PhoA activity. The plates were incubated at 37°C for 2 days before colonies were picked.

For the second series, a 100- $\mu$ l sample of cells was infected with 100- $\mu$ l of phage lysate for 20 min at 37°C and directly plated on either (1) glycerol-E medium containing 50 mM CaCl<sub>2</sub>, 30  $\mu$ g/ml kanamycin and 40  $\mu$ g/ml XP or (2) glycerol-E medium containing 5 mM CaCl<sub>2</sub>, 180  $\mu$ g/ml kanamycin and 40  $\mu$ g/ml XP. These plates were also incubated at 37°C for 2 days before colonies were picked.

Atypically small colonies on the plates were judged to be potential calcium-sensitive mutants. Those colonies that grew on glycerol-E medium but not on glycerol-E medium containing 50 mM CaCl<sub>2</sub> (thus indicating sensitivity to calcium) were purified and retained.

### Calcium-sensitivity assay

Calcium-sensitivity was later defined as the inability to grow in the presence of 75 mM CaCl<sub>2</sub>. Calcium-sensitivity was measured by two different growth inhibition assays. Initially, calcium-sensitivity was measured by growth inhibition assay of Brey and Rosen<sup>23</sup>. Cells were screened for their ability to grow on E medium plates supplemented with 0 mM, 25 mM, 50 mM, 75 mM, or 100 mM CaCl<sub>2</sub>. The plates were incubated at 37°C and the growth

results were monitored at 24 h and 48 h. After 48 h, calcium-sensitive cells are unable to grow on E medium plates supplemented with 75 mM CaCl<sub>2</sub>. Growth inhibition was also determined from turbidity measurements in broth cultures. During this assay, cells were inoculated in E medium broth supplemented with glycerol and different concentrations (0 mM, 25 mM, 50 mM, 75 mM, or 100 mM) of CaCl<sub>2</sub>, and were incubated at 37°C for 8 h. The optical density at a wavelength of 600 nm was measured at 0 and 8 h.

#### Cation sensitivity assay

The sensitivity to various other cations was monitored by growth inhibition assay of the calcium-sensitive cells as described by Brey and Rosen<sup>23</sup>. The minimum inhibitory concentration (MIC) values were determined for each cation tested. MIC is the lowest concentration of cation at which all bacterial growth is stopped.

#### Chemotaxis assay

Chemotactic ability was measured by swim-ring migration assay<sup>24</sup>. Tryptone swarm plates containing 1% bacto-tryptone, 0.5% NaCl, and 0.25% bactoagar (Difco Laboratories, Franklin Lakes, New Jersey) were inoculated with a stab of approximately 10<sup>6</sup> cells at the centre of the plates and incubated at 30°C for 8 to 14 h. The ring diameters were measured at the end of the experiment.

#### Analysis of free-swimming behaviour

Bacterial swimming behaviour was observed at 30°C by phase-contrast microscopy at a magnification of 400x. The cells in these behavioural assays were suspended in chemotaxis medium (10 mM potassium phosphate pH 7.0, 0.1 mM potassium EDTA, and 1 mM L-methionine) or in filtered spent/used growth medium to an optical density at 590 nm of 0.1. The microscopic behaviour was videotaped and analyzed by computer<sup>25</sup>.

#### Alkaline phosphatase assay

Alkaline phosphatase activity was measured using both solid (for all mutants) and liquid media (only for calcium-sensitive transposon mutant Pho43). Cells were grown at 37°C in LB medium containing appropriate antibiotics. Overnight cultures were then streaked for isolation on E medium plates containing the chromogenic alkaline phosphatase substrate 5-bromo-4-chloro-3-indolyl phosphate (XP). The parental strain CC118 lacks

functional alkaline phosphatase. Only colonies that produced alkaline phosphatase fusion on the outside of the cell or in the periplasmic space would turn blue. Following overnight growth at 37°C, successful calcium-sensitive mutants with functional alkaline phosphatase were monitored for blue color production.

#### DAPI experiments

Prior to staining the cells were first fixed with toluene and treated with chloramphenicol to condense their genomes. The fixed cells were incubated with the fluorescent DNA binding dye DAPI (4',6'-diamidino-2-phenylindole 2 HCl), and observed by the use of phase contrast microscopy and fluorescence microscopy at a total magnification of 1,000x.

#### Arbitrarily primed PCR

To identify the *cal* gene sequence, the region of the genes fused to the reporter gene (*phoA*) were amplified by PCR and sequenced. To amplify partial regions of *cal* genes fused to the transposon, we used the arbitrarily-primed PCR method<sup>26-27</sup>. This method involved two rounds of PCR amplification using arbitrary primers to prime from the chromosome and primers specific to *TnphoA*. During the first round of PCR, the primer (Tn-R) with sequence homology to the right end of the transposon *TnphoA* and arbitrary primer ARB1 were used. The PCR parameters for first round were as follows: (1) 95°C 5 min, (2) 6 cycles of 95°C for 30 s, 30°C for 30 s, 72°C for 1.5 min, (3) 30 cycles of 95°C for 30 s, 45°C for 30 s, and 72°C for 2 min. During the second round of PCR, a 5 μl aliquot of first round PCR product was used as template and the primers ARB2 and Tn-I were used for amplification. The PCR parameters for second round were as follows: 30 cycles of 95°C for 30 s, 45°C for 30 s, 72°C for 2 min. The final PCR products were purified from an agarose gel with the aid of  $\lambda$ -agarase. The primers (ARB1, ARB2, Tn-I and Tn-R) used in this experiment are listed in Table 2. These PCR amplified products were sequenced using primers (*TnphoA*-int and Tn-R) located close to the fusion site on *TnphoA*. DNA sequence data were analyzed by sequence analysis software DNASTar for Mac. The resultant edited sequences, ORFs present in the edited sequences, as well as translated sequences were used to query the *E. coli* genomic database<sup>28</sup>.

#### Bacterial growth conditions for DNA macroarray experiments

To provide a comprehensive insight into calcium homeostasis, the global expression profiles of wild type *E. coli* MG 1655 were

**Table 2.** Primers used in arbitrarily-primed PCR and sequencing reaction

Primer	Primer sequence (5' → 3')	Reference
ARB1	GGCCACGCGTCGACTAGTACNNNNNNNNNNGATAT	This study
ARB2	GGCCACGCGTCGACTAGTAC	This study
Tn-I	CCTTCGGCATAATTACGTGC	This study
Tn-R	GCAGTCTGATCACCCGTTAAA	This study
TnphoA-int	TTTCCAGAACAGGGCAAAC	This study
TnphoA-F	TGCAAGTTGAAGGTGCGTCAATCG	This study
TnphoA-R	ATGAGATGCCCTGCAAGCAATTCG	This study

determined under three different conditions: (1) growth medium containing elevated levels of calcium (the addition 75 mM CaCl<sub>2</sub>), (2) growth medium containing depleted levels of calcium (the addition of 10 mM EGTA), and (3) growth medium control (untreated). Overnight cultures of *E. coli* MG 1655 were used to inoculate fresh E media and the cultures were incubated at 37°C with rotary oration. For the array experiments cultures were grown to an OD<sub>600</sub> of 0.4 to 0.5 and then either 75 mM calcium or 10 mM EDTA was added. After 30 min of rotary aeration at 37°C, total RNA was quickly extracted by using QiagenRNeasy kit (Qiagen, Inc., Valencia, CA). Proper precautions were taken to avoid contamination with RNases and a non-denaturing electrophoresis gel was run with the extracted RNA to check the integrity of RNA.

#### Synthesis of <sup>33</sup>P-labeled cDNA probe

Sigma-Genosys Biotechnologies, Inc. had developed commercially available cDNA primers that are specifically designed to preferentially label cDNA from mRNA. These C-terminal primer sets (4,290 ORF-specific C-terminal primers) were used to generate the hybridization probe according to the manufacturers recommendations. As recommended by the manufacturer, <sup>33</sup>P- $\alpha$ -dCTP was used to label the cDNA. The <sup>33</sup>P-labeled cDNA was purified and unincorporated-labeled nucleotides were removed by the use of Sephadex G-25 gel-filtration spin columns. An estimation of percentage incorporation of <sup>33</sup>P-dCTP into the cDNA was determined by using a hand-held Geiger counter or by scintillation counts of samples before and after column purification. The labeled cDNA probe was then ready to be used in a hybridization reaction with the Panorama *E. coli* Gene Arrays (Sigma-Genosys Biotechnologies, Inc.).

#### Hybridization with panorama *E. coli* gene array

Following its preparation and purification, the <sup>33</sup>P-labeled cDNA was hybridized to the Panorama gene array by the use of roller bottles in a hybridization oven. The hybridization and washing steps were performed according to the protocol specified in Panorama *E. coli* cDNA Labeling and Hybridization Kit. The DNA array blots were rinsed in 2X SSPE (1X SSPE: 0.18 M NaCl, 10 mM NaH<sub>2</sub>PO<sub>4</sub>, 1 mM EDTA, pH 7.7) and pre-hybridized in pre-warmed hybridization solution (5X SSPE, 2% SDS, 1X Denhardt's reagent, 100  $\mu$ g/ml sheared salmon sperm DNA) at 65°C for 1 hr at 6 rpm. The entire <sup>33</sup>P-labeled cDNA was first denatured at 95°C for 10 min and then added to 3 ml of hybridization buffer and the blot were hybridized in this solution for 15 h at 65°C. After overnight incubation, the blots were washed with buffer (0.5X SSPE, 0.2% SDS) three times at room temperature. The blots were then washed three more times with pre-warmed (65°C) buffer in the hybridization oven at 65°C for 20 min at 6 rpm. Washed blots were finally air-dried briefly for 5 min and wrapped in clear plastic food wrap and exposed to a phosphorimager screen (Bio-Rad Laboratories) for 24 h.

#### Data acquisition by phosphorimaging and analysis of the arrays

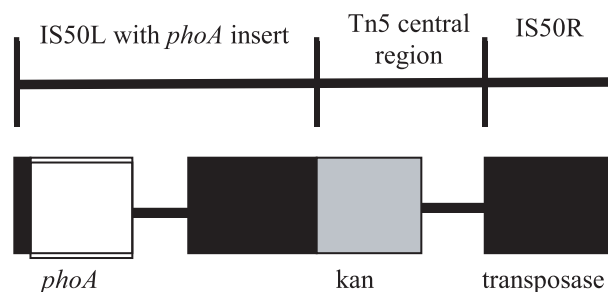
The gene expression signals were measured by the use of a Bio-Rad PhosphorImager with a Kodak Low Energy Storage Phosphor Screen. Typically, the arrays were exposed overnight to yield quantifiable results. For quantification, imaging screens were

scanned at a 50 $\mu$ m pixel size and analyzed by Quantity One software (Bio-Rad Laboratories, Hercules, CA) based on the spot coordinates. Each gene has two corresponding spots and the pixel density (intensity) of each spot representing each gene in the array was measured and corrected for the background. The average signal of the pair of duplicate spots were subsequently determined and normalized for comparative studies. To standardize the data from experiments the average intensity for each spot was expressed as a percentage of the total of intensities for all of the spots on the DNA array.

## Results and Discussion

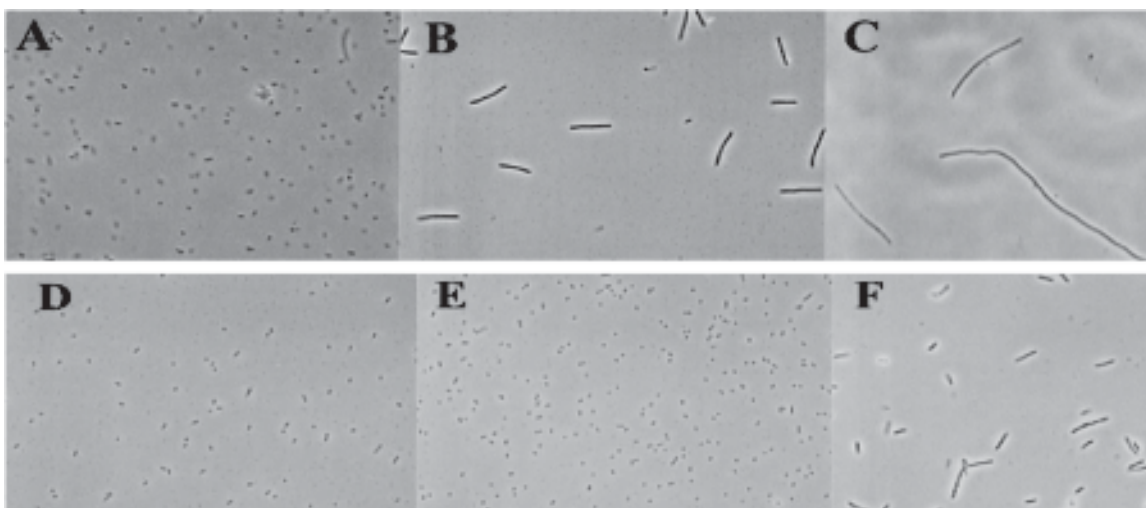
### Isolation of calcium-sensitive mutants of *E. coli* by *TnphoA* mutagenesis

The 7.7 kb transposon *TnphoA* contains Kan<sup>r</sup> gene as a selective marker and the reporter gene for alkaline phosphatase *phoA* (Figure 1). The transposon, *TnphoA*, used as a mutagenic agent for generation and isolation of new calcium-sensitive mutants. The reporter gene for alkaline phosphatase *phoA* has no promoter or Shine Dalgarno sequences, and is only expressed when the fusion is in-frame. When inserted in frame and in proper orientation, *TnphoA* will randomly mutagenize bacterial chromosome by disrupting the gene the transposon will insert itself in. In addition, *TnphoA* will fuse alkaline phosphatase to the amino terminal of the protein product of the disrupted gene. Alkaline phosphatase is functional when it is fused to a signal that promotes the export of the protein to the envelope of the cell (such as periplasmic, outer membrane or cytoplasmic membrane proteins). Our search for calcium sensitive *E. coli* mutants was targeted to membrane protein genes by essentially isolating *phoA* gene fusions. Infection of a *phoA* deletion strain CC118 with  $\phi$ *TnphoA*, followed by penicillin enrichment and subsequent screening of blueness of colonies on XP resulted in mutants with *TnphoA* insertions exhibiting PhoA activity.



**Figure 1.** The transposon *TnphoA* (Manoil and Beckwith<sup>18</sup>). The 7.7 Kbp transposon is derived from *Tn5* with the selective marker *Kan* and the reporter gene *phoA*. Successful (in frame) gene-transposon fusion allows for identification of secreted and transmembrane proteins.

Following transposon mutagenesis, penicillin enrichment was used to aid in the isolation of calcium-sensitive mutants. Several new calcium-sensitive mutants were identified by this transposon mutagenesis procedure. The calcium-sensitive mutants generated are described in Table 1 and photomicrographs of some mutants are included in Figure 2. The mutants fell into three



**Figure 2.** Photomicrograph of wild type *Escherichia coli* and calcium-sensitive mutants. (A) CC118 with wild type morphology, (B) Cal102 cells with a snake-like morphology consist of up to 10 times the length of the wild type CC118, (C) Cal134 cells were more elongated at about 10 to 20 times the length of CC118, (D) and (E) Cal 117 and Pho43, respectively, had wild type morphology, and (F) Pho43 are long like Cal 102.

general classes: (i)  $\text{Ca}^{2+}$ -sensitive chemotaxis mutants, (ii)  $\text{Ca}^{2+}$ -sensitive cell division mutants, and (iii)  $\text{Ca}^{2+}$ -sensitive mutants that showed no defects in cell division or chemotaxis (Table 3). While some of the mutants were sensitive to 50 mM  $\text{CaCl}_2$ , sensitivity measured by growth inhibition assay revealed the inability of most mutants to grow in the presence of 75 mM  $\text{CaCl}_2$  on plates or in broth cultures.

#### *Physiological and Biochemical Properties of Calcium-Sensitive Mutants*

The cell division mutants (e.g., Cal134) were easily identified microscopically as long “snake-like” cells (Figure 2). One of the  $\text{Ca}^{2+}$ -sensitive cell division mutants (Cal134) grew as a long filamentous cell that was over 20 cell-body-length in size.

Multiple chromosomes within these snake-like cells were observed following DAPI staining of this mutant (Figure 3). These results

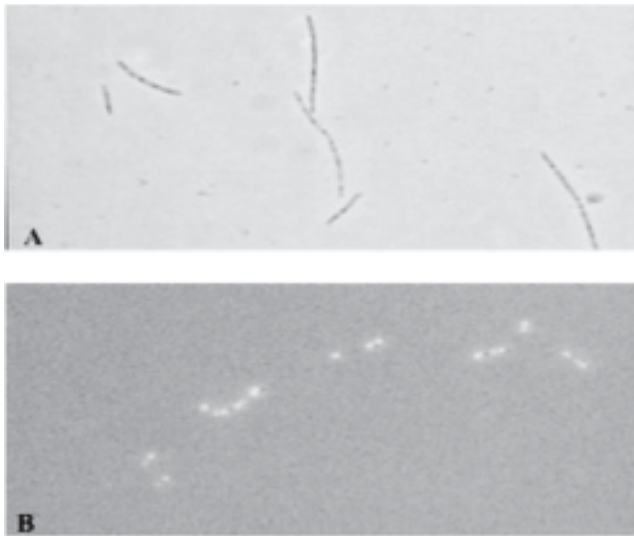
imply that this mutant was defective in cell division, but not DNA replication.

The generation of these  $\text{Ca}^{2+}$ -sensitive cell division mutants was not unexpected. Holland and co-workers<sup>29-30</sup> isolated mutants that were resistant to different calcium antagonists and are defective in cell division. They proposed a model suggesting that calcium plays a role in the control of bacterial growth and its cell cycle<sup>31-32</sup>. The sensitivity of these calcium-sensitive mutants to other cations was tested and their minimal inhibitory concentration (MIC) values for these cations are shown in Figure 4. Besides calcium-sensitivity, all of the mutants exhibited increased sensitivities to several divalent cations ( $\text{Ni}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Cu}^{2+}$ , and  $\text{Cd}^{2+}$ ). Mutant Cal117 was sensitive to all of the cations tested. Mutants Cal134 and Pho43 exhibited sensitivity to  $\text{Co}^{2+}$  and  $\text{Zn}^{2+}$ , while Cal134 was also sensitive to  $\text{Cu}^{2+}$ . Cal102 was sensitive to all of the cations except  $\text{Ni}^{2+}$ , and

**Table 3.** Properties of calcium-sensitive mutants

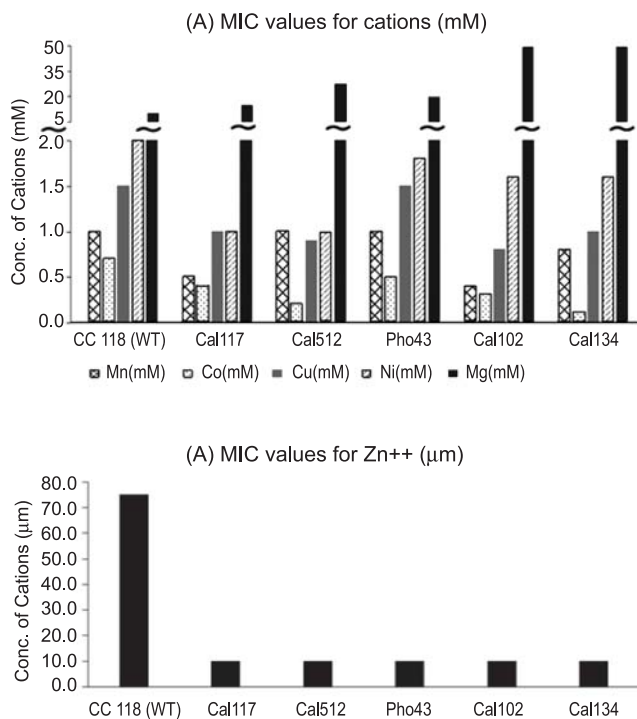
Strain	Morphology	Motility	Swim rate mm per 8 hours	Alkaline phosphatase (plate assay)
CC118 (WT)	Short rods	Smooth-tumbly	49	–
Cal117	Medium snakes	Smooth	0	++
Cal512	Medium snakes	Smooth	2	+
Cal102	Medium snakes	Tumbly	14	++
Pho43	Sausage snakes	Non-motile	0	++
Cal134	Long snakes	Smooth	4	++
PhoA4	Short rods	Smooth-tumbly	37	+++
PhoC23	Short rods	Tumbly	23	–
Cal 719	Medium snakes	Smooth-tumbly	31	ND
Cal526	ND	ND	11	ND
Cal504	ND	ND	12	ND

Medium = 5-8 cell lengths; Long = 10-20 cell lengths; – = No color; + = Light blue; ++ = Medium blue; +++ = Dark blue; ND = Not determined.



**Figure 3.** The *Cal134* mutant is defective in cell division, but not DNA replication. DAPI stained phase contrast (A) and fluorescent (B) pictures of *Cal134* long snake cell division mutants.

*Cal512* exhibited a similar pattern of sensitivity as *Cal117* though it was not sensitive to  $Mn^{2+}$ . It is noteworthy to mention that all calcium-sensitive mutants were resistant to  $Mg^{2+}$  as shown in Figure 4A.



**Figure 4.** *Ca*-sensitive mutants were sensitive to other divalent cations. MIC values for cations tested were determined as described in the Methods. Mutants *Cal117* and *Cal512* exhibit a similar pattern of cation sensitivity (except for  $Mg$ ), while *Cal102* and *Cal134* exhibit similar profiles. The wild type *CC118* and mutant *Pho43* have identical MICs for  $Mn^{2+}$  and  $Cu^{2+}$ .

In other ion transport systems, mutants defective in ion transport show increased sensitivity to diverse additional ions.  $Na^+$  transport mutants are resistant to  $Li^+$ <sup>33</sup>, while  $Mg^{2+}$  transport (*corA*) mutants are resistant to  $Co^{2+}$  and sensitive to  $Ca^{2+}$ <sup>34</sup>. We expected that the *cal* mutants isolated above would show sensitivities to other cations or that they would show an increased resistance to some cations.

Table 3 summarizes the physiological and biochemical characterization data for a comprehensive representation of observations made on the transposon mutants.

#### Molecular characterization of calcium-sensitive mutants

The sites of the gene fusions for three mutants were identified by arbitrarily-primed PCR (arbPCR). Since the PCR products of arbPCR contained part of the upstream regions of the transposon *TnphoA* fused to the disrupted gene, the start of *cal* gene was easily recognized. The resulting partial gene sequence was used to search the compiled DNA sequence databank of *E. coli* genome by the use of the BLAST program at NCBI. The sites of the gene fusion for three *cal* mutants were at the *fdoG*, *gpt* and *pqi* genes (Table 4). The *fdoG* gene codes for the  $\alpha$ -subunit of formate dehydrogenase<sup>35</sup>, while the *gpt* gene codes for guanine-xanthine phosphoribosyl transferase phosphotransferase<sup>36</sup> and *pqi* gene is induced by paraquat and regulated by SoxRS<sup>37</sup>. The arbitrarily-primed PCR fragment of *Cal134* exhibited 96% sequence homology with *fdoG* gene and also a significant match as evidenced by the *e* value of  $5e-12$ . Similarly, the *cal* genes of *PhoC23* and *PhoA4* showed significant matches to genes *gpt* and *pqiB* (*e* value of  $2e-44$  and  $1e-25$ , respectively) with 98% sequence identity for both.

The site of the *TnphoA* insertion was detected by hybridization of a DIG-labeled *TnphoA* derived probe to membrane blotted nucleic acids. Hybridization experiments using this *TnphoA* derived probe confirmed that each mutation was distinct and the result of the single insertion of *TnphoA* (data not shown). The site of insertion was also confirmed by PCR (data not shown).

The sites of the gene fusion for three *cal* mutants were the following: gene (*Cal134*), *gpt* gene (*PhoC23*), *pqi* gene (*PhoA4*). Our initial observations with the *TnphoA*-generated *cal* mutants suggest that their calcium-sensitivity was the result of a number of diverse genes being disrupted by the insertion of the transposon. These results suggest a pleiotropic nature for the *cal* mutations and indicate that many genes may be regulated by calcium. Because of the results from these experiments, we wanted to investigate global gene expression under growth conditions with and without calcium with a broader goal of identifying all of the calcium-regulated genes in *E. coli*.

#### Panorama *E. coli* gene array

Panorama *E. coli* DNA macroarrays (Sigma\_Genosys) representing all 4,290 protein-coding genes were used to identify those genes that were globally regulated by calcium and to observe the effects

**Table 4.** *cal* gene mutations identified by ARB-PCR

Mutant	<i>cal</i> gene	Gene function	e-value	% identity
Cal134	<i>fdoG</i>	Formate dehydrogenase (Allows the use of formate as major electron donor during aerobic respiration)	5e-12	96
PhoC23	<i>gpt</i>	Guanine-hypoxanthine phosphoribosyltransferase (Involved in salvage of nucleosides and nucleotides)	2e-44	98
PhoA4	<i>pqiB</i>	Paraquat-inducible protein (Inducible by superoxide generating radical paraquat and regulated by SoxRS)	1e-25	98

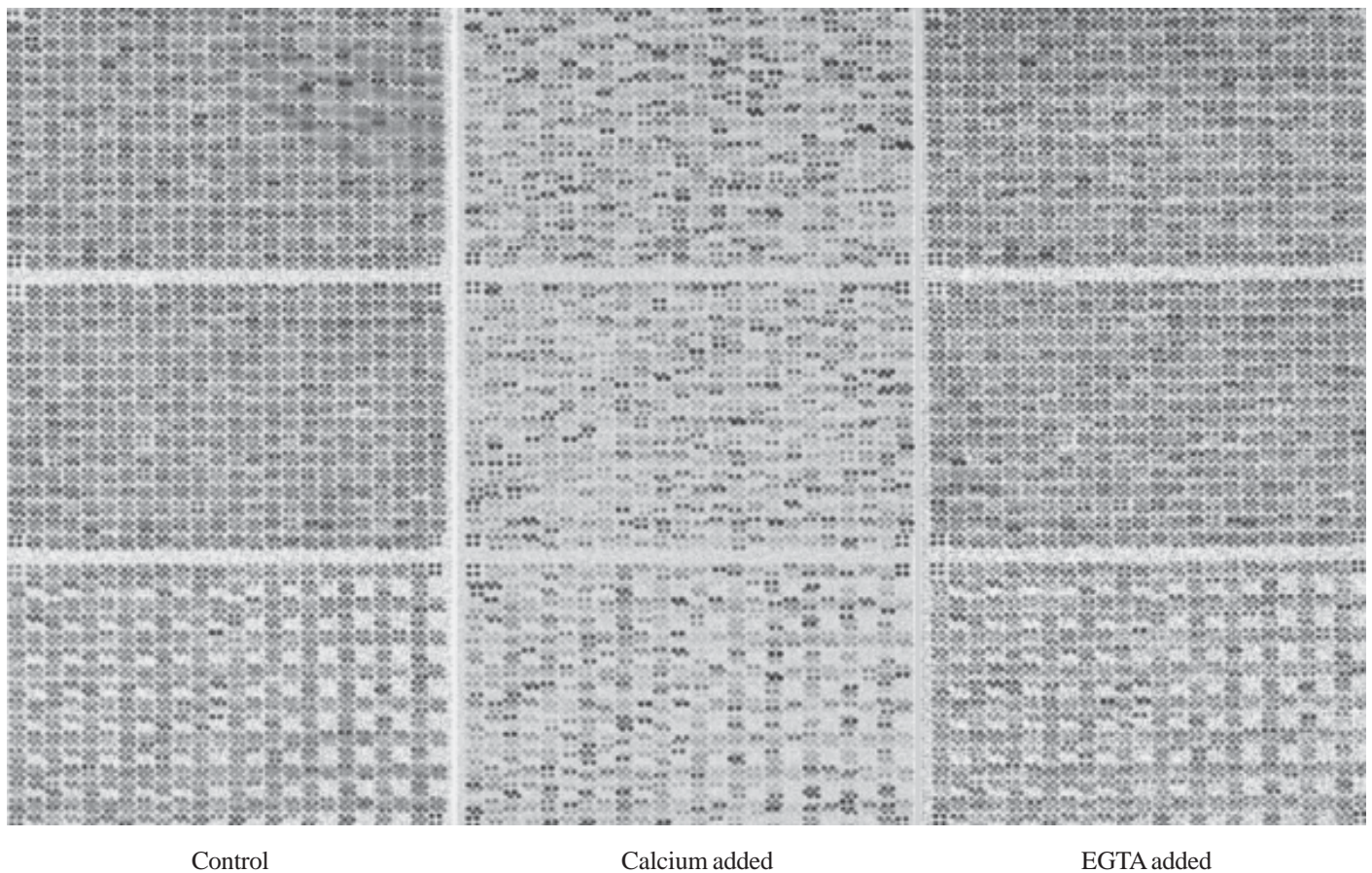
of elevated (75 mM Ca) and depleted calcium (10 mM EGTA) levels on the global transcription profile (Figure 5).

The 23S and 16S ribosomal RNA bands were clearly visible at about 2:1 ratio (23S:16S) of staining intensity. The processed RNA samples were devoid of any genomic DNA contamination as evidenced by agarose gel electrophoresis.

The corresponding spots from samples on arrays were compared to identify the fold-induction (up-regulation) or fold-reduction

(down-regulation) in expression between the samples (Figure 5). By this method of array analysis a 2-fold difference in expression was considered as important. Changes in expression (fold change) of genes during elevated and depleted calcium conditions are shown in Table 5.

The findings from our study of calcium homeostasis represent a dynamic state with hitherto unknown regulatory activity throughout *E. coli* genome. It is very likely that diverse genes are under calcium regulation as evidenced by changes in gene



**Figure 5.** Panorama *E. coli* gene array exhibiting global gene expression profiles of *E. coli* MG1655 following growth on *E* medium (control), *E* medium supplemented with 75 mM calcium, and *E* medium supplemented with 10 mM EGTA.



expression in elevated or depleted calcium. The next logical extension of our study will include an investigation into quantitative analysis of gene expression of subsets of genes in wild-type and mutant strains with addition of calcium and chelation of calcium.

### Acknowledgements

This investigation was supported in part by Grant-in-Aid 96077285 from The American Heart Association and by the College of Life Science and Agriculture, The University of New Hampshire. JH was supported by a Summer Undergraduate Research Fellowship from The University of New Hampshire.

### References

- Gangola P and Rosen BP. 1987. Maintenance of intracellular calcium in *Escherichia coli*. *J. Biol. Chem.* **262**: 12570-12574.
- Tisa LS and Adler J. 1992. Calcium ions are involved in *Escherichia coli* chemotaxis. *Proc Natl Acad Sci USA.* **89**: 11804-11808.
- Tisa LS and Adler J. 1995a. Chemotactic properties of *Escherichia coli* mutants having abnormal Ca<sup>2+</sup> content. *J. Bacteriol.* **177**(24): 7112-7118.
- Tisa LS and Adler J. 1995b. Cytoplasmic free-Ca<sup>2+</sup> level rises with repellents and falls with attractants in *Escherichia coli* chemotaxis. *Proc Natl Acad Sci USA.* **92**: 10777-10781.
- Tisa LS, Olivera B and Adler J. 1993. Inhibition of *Escherichia coli* chemotaxis by ù-conotoxin, a calcium channel blocker. *J Bacteriol.* **175**: 1235-1238.
- Tisa LS, Sekelsky JJ and Adler J. 2000. Effects of organic antagonists of Ca<sup>2+</sup>, Na<sup>+</sup>, and K<sup>+</sup> on chemotaxis and motility of *Escherichia coli*. *J Bacteriol.* **182**: 4856-4861.
- Watkins NJ, Knight MR, Trewavas AJ and Campbell AK. 1995. Free calcium transients in chemotactic and non-chemotactic strains of *Escherichia coli* determined by using recombinant aequorin. *Biochem J.* **306**: 865-869.
- Nakashima K, Sigiua A, Momoi H and Mizuno T. 1992. Phosphotransfer signal transduction between two regulatory factors involved in the osmoregulated *Kdp* operon of *Escherichia coli*. *Mol Microbiol.* **6**: 1774-1784.
- Rampersaud A, Utsumi R, Forst SA and Inouye M. 1991. Ca<sup>2+</sup>-enhanced phosphorylation of a chimeric protein kinase involved with bacterial signal transduction *J Biol Chem.* **266**: 7633-7637
- Sweeney ST, Freestone P and Norris V. 1995. Characterization of eukaryotic-like kinase activity in *Escherichia coli* using the gene-protein database. *FEMS Microbiol Lett.* **127**: 133-138.
- Sandler N and Keynen A. 1992. Membrane-protein phosphorylation in *Bacillus subtilis* cell cycle. *FEMS Microbiol Letts.* **95**: 241-246.
- Stowe DJ, Atkinson T and Mann NH. 1989. Protein kinase activities in cell-free extracts of *Streptomyces coelicolor* A3(2). *Biochimie.* **71**: 1101-1105.
- Onek LA and Smith RJ. 1992. Calmodulin and calcium mediated regulation in prokaryotes. *J Gen Microbiol.* **138**: 1039-1049.
- Laoudi D, Anderson CL, Bras A, Glodberg M, Jacq A and Holland IB. 1994. EGTA induces the synthesis of in *Escherichia coli* of three proteins that cross-react with calmodulin antibodies. *Mol Microbiol.* **13**: 445-457.
- Brey RN and Rosen BP. 1979. Properties of *Escherichia coli* mutants altered in calcium/proton antiport activity. *J Bacteriol.* **139**: 824-834.
- Ivey DM, Guffanti AA, Zemsky J, Pinner E, Karpel R, Padan E, Schuldiner S and Krulwich TA. 1993. Cloning and characterization of a putative Ca<sup>2+</sup>/H<sup>+</sup> antiporter gene from *Escherichia coli* upon functional complementation of Na<sup>+</sup>/H<sup>+</sup> antiporter-deficient strains by the overexpressed gene. *J Biol Chem.* **268**: 11296-11303.
- Naseem R, Wann KT, Holland IB and Campbell AK. 2009. ATP regulates calcium efflux and growth in *E. coli*. *J Mol Biol.* **391**: 42-56.
- Manoil C and Beckwith J. 1985. TnpHoA: A transposon probe for protein export signals. *Proc Natl Acad Sci USA.* **82**: 8129-8133.
- Blattner FR, Plunkett III G, Bloch CA, Perna NT, Burland V, Riley M, Collado-Vides J, Glasner JD, Rode CK, Mayhew GF, Gregor J, Davis NW, Krikpatrick HA, Goeden MA, Rose DJ, Mau B and Shao Y. 1997. The complete genome sequence of *Escherichia coli* K-12. *Science.* **277**: 1453-1474.
- Koh YS and Roe JH. 1995. Isolation of a novel paraquat-inducible (*pqi*) gene regulated by the *soxRS* locus in *Escherichia coli*. *J Bacteriol.* **177**(10): 2673-2678.
- Brey RN, Beck JC and Rosen BP. 1978. Cation/proton antiport systems in *Escherichia coli*. *Biochem Biophys Res Commun.* **83**: 1588-1594.
- Vogel HJ and Bonner DM. 1956. Acetylornithinase of *Escherichia coli*: Partial purification and some properties. *J Biol Chem.* **218**: 97-106.
- Brey RN and Rosen BP. 1979. Properties of *Escherichia coli* mutants altered in calcium/proton antiport activity. *J Bacteriol.* **139**: 824-834.
- Adler J. 1966. Effect of amino acids and oxygen on chemotaxis in *Escherichia coli*. *J Bacteriol.* **92**(1): 121-129.
- Sager B, Sekelsky JJ, Matsumura P and Adler J. 1988. Use of a computer to assay motility in bacteria. *Anal Biochem.* **173**: 271-277.
- Caetano-Annoles G. 1993. Amplifying DNA with arbitrary oligonucleotide primers. *PCR Methods Appl.* **3**: 85-92.
- O'Toole GA and Kolter R. 1998. Initiation of biofilm formation in *Pseudomonas fluorescens* WCS365 proceeds via multiple, convergent signalling pathways: A genetic analysis. *Mol Microbiol.* **28**: 449-461.
- Altschul SF, Gish W, Miller W, Myers EW and Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol.* **215**: 403-410.
- Bouquin N, Tempête M, Holland IB and Séror S. 1995. Resistance to trifluoroperazine, a calmodulin inhibitor, maps to the *fabD* locus in *Escherichia coli*. *Mol Gen Genet.* **246**: 628-637.
- Casaregola S, Chen M, Bouquin N, Norris V, Jacq A, Goldberg M, Margaron S, Tempête M, McKenna S, Sweetman H, Bernard S, McGurk G, Séror SJ and Holland IB. 1991. Analysis of a myosin-like protein and the role of calcium in the *Escherichia coli* cell cycle. *Res Microbiol.* **142**: 201-207.
- Holland IB, Casaregola S and Norris V. 1990. Cytoskeletal elements and calcium: Do they play a role in the *Escherichia coli* cell cycle. *Res Microbiol.* **142**: 131-137.
- Norris V, Seror SJ, Casaregola S and Holland IB. 1988. A single calcium flux triggers chromosome replication, segregation and septation in bacteria: A model. *J Ther Biol.* **134**: 341-350.
- Park MH, Wong BB and Lusk JE. 1976. Mutants in three genes affecting transport of magnesium in *Escherichia coli*: Genetics and physiology. *J Bacteriol.* **126**: 1096-1103.
- Maguire ME. 1993. Genetics and molecular biology of magnesium transport in *Salmonella typhimurium*. In *Magnesium and the Cell* (Birch NJ ed), pp 157-178. Academic Press, New York.
- Wang H and Gunsalus RP. 2003. Coordinate Regulation of the *Escherichia coli* formate dehydrogenase *fdnGHI* and *fdhF* genes in response to nitrate, nitrite, and formate: Roles for NarL and NarP. *J Bacteriol.* **185**(17): 5076-5085.
- Pratt D and Subramani S. 1983. Nucleotide sequence of the *Escherichia coli* xanthine-guanine phosphoribosyltransferase gene. *Nucl Acids Res.* **11**: 8817-8823.
- Koh YS and Roe JH. 1995. Isolation of a novel paraquat-inducible (*pqi*) gene regulated by the *soxRS* locus in *Escherichia coli*. *J Bacteriol.* **177**(10): 2673-2678.