Genetic disruption of the core circadian clock impairs hippocampus-dependent memory

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Perturbing the circadian system by electrolytically lesioning the suprachiasmatic nucleus (SCN) or varying the environmental light/dark schedule impairs memory, suggesting that memory depends on the circadian system. We used a genetic approach to evaluate the role of the molecular clock in memory. Bmal1−/− mice, which are arrhythmic under constant conditions, were examined for hippocampus-dependent memory, LTP at the Schaffer-collateral synapse, and signal transduction activity in the hippocampus. Bmal1−/− mice exhibit impaired contextual fear and spatial memory. Furthermore, LTP in hippocampal slices from Bmal1−/− mice is also significantly decreased relative to that from wild-type mice. Activation of Erk1,2 MAP kinase (MAPK) during training for contextual fear memory and diurnal oscillation of MAPK activity and cAMP in the hippocampus is also lost in Bmal1−/− mice, suggesting that the memory defects are due to reduction of the memory consolidation pathway in the hippocampus. We conclude that critical signaling events in the hippocampus required for memory depend on BMAL1.

Hippocampus-dependent learning and memory is initiated by a calcium influx mediated through NMDA receptors (Morris et al. 1986; Tsien et al. 1996) and leads to the activation of several signaling pathways, including the CAMP and Erk1,2 MAP kinase (MAPK) pathways (Adams and Sweatt 2002; Wang and Storm 2003). Activation of CaM-stimulated adenyl cyclases and MAPK causes stimulation of CRE-mediated transcription (Impney et al. 1998; Athos et al. 2002; Sindreu et al. 2007). Perturbing any of these signaling events disrupts memory consolidation (Bourchuladze et al. 1994; Atkins et al. 1998; Wong et al. 1999; Athos et al. 2002; Pittenger et al. 2002).

Despite the potential for memories to persist over the lifetime of an individual, the proteins that underlie strengthened synaptic connections degrade. Recurring rounds of transcription and protein synthesis may contribute to the persistence of memories (Bekinschtein et al. 2011; Eckel-Mahan et al. 2008). Specifically, Eckel-Mahan et al. (2008) showed that the CAMP/MAPK/CREB transcriptional pathway undertakes a circadian oscillation in the hippocampus that, when blocked, inhibits the maintenance of contextual fear memory in mice. The mammalian circadian system may provide the signal to coordinate returning rounds of transcription important for memory maintenance.

Previous research has established that time of day can influence learning and memory of fear conditioning under both light/dark entrained and free-running conditions (Chaudhury and Colwell 2002; Eckel-Mahan et al. 2008). Hippocampal LTP also depends on time of day under light/dark entrained conditions (Chaudhury et al. 2005). This time-of-day influence on memory and LTP suggests involvement of the circadian system. Perturbing the circadian system through environmental disruption of lighting conditions or lesioning the master circadian regulator can impair memory (Tapp and Holloway 1981; Devan et al. 2001; Lyons et al. 2006; Ruby et al. 2008; Loh et al. 2010; Phan et al. 2011). The suprachiasmatic nucleus (SCN) is the master regulatory clock in mammals. The SCN receives light input from the environment via the retinohypothalamic pathway and synchronizes peripheral oscillators via synaptic connection or diffusible factors (Antle and Silver 2005). A molecular clock comprised of positively and negatively regulated transcriptional/translational loops drives the oscillations in central and peripheral sites with a period of about 24 h (Ko and Takahashi 2006). BMAL1 and CRYPTOCHROME dimerize and bind to E-box elements to promote the transcription of circadian-regulated genes, including the heterodimer’s negative regulators, Period and Cryptochrome. PERIOD and CRYPTOCHROME dimerize and translocate to the nucleus to inhibit the activity of the BMAL1:CRYPTOCHROME heterodimer. A second loop involving the competitive binding of REV-ERBα and RORα at RORE sites at BMAL1 promoter regulates BMAL1 level.

Only the Bmal1−/− mouse is completely arrhythmic in constant conditions and exhibits impaired entrainment to a light/dark cycle (Bunger et al. 2000). Other molecular clock knockout mouse strains retain some rhythmicity due to compensatory functional isoforms and present with subtle or no memory deficits (Garcia et al. 2000; Debruyne et al. 2006; Zueger et al. 2006; Van der Zee et al. 2008). Bmal1−/− mice provide an intriguing model to genetically address the role of the molecular clock in memory.

We hypothesize that the molecular clock is required for the diurnal oscillation of signaling activity in the hippocampus and hippocampus-dependent memory. Here, we report that Bmal1−/− mice have impaired contextual fear memory, defects in learning and spatial memory, impaired LTP at the Schaffer-collateral synapse, as well as no diurnal change in cAMP and MAPK activity.
Results

*Bmal1<sup>−/−</sup>* mice exhibit impaired hippocampus-dependent learning and memory

Consistent with previous reports (Bunger et al. 2000), *Bmal1*<sup>−/−</sup> mice are arrhythmic under dark-dark (DD) conditions while *Bmal1*<sup>+/+</sup> littermates exhibit an expected τ close to 24 h (Fig. 1C,D). *Bmal1*<sup>−/−</sup> mice also exhibit difficulties entraining to a 12-h light-dark (LD) cycle (Fig. 1A,B), thus all experiments were performed under diurnal conditions.

*Bmal1*<sup>+/+</sup> and *Bmal1*<sup>−/−</sup> mice were trained and tested for contextual fear conditioning from ZT3 (3 h after light onset) to ZT5, the peak of MAPK activity and fear memory (Chaudhury et al. 2002; Eckel-Mahan et al. 2008), and under red light from ZT15 to ZT17, the trough of MAPK activity and fear memory. Mice explored a novel context for 2 min before receiving a 0.7-mA foot shock. *Bmal1*<sup>−/−</sup> mice froze significantly less than their wild-type littermates when they were tested 24 h after training (*F*<sub>5,45</sub> = 31.44, *P* < 0.0001, one-way ANOVA, Tukey post-hoc test) (Fig. 2A). While *Bmal1*<sup>−/−</sup> mice did learn to associate the novel context with a foot shock (*P* < 0.0001 and *P* < 0.05, Tukey post-hoc test) (Fig. 2A), their long-term memory for contextual fear conditioning was not as strong as that of *Bmal1*<sup>+/+</sup> controls. *Bmal1*<sup>−/−</sup> mice contextual fear memory was comparable to that of *Bmal1*<sup>+/+</sup> controls when trained and tested during the night (Fig. 2B).

In the Morris water maze, *Bmal1*<sup>+/+</sup> and *Bmal1*<sup>−/−</sup> mice were trained to navigate to a hidden platform for 8 d from ZT4 to ZT8. Throughout this training, *Bmal1*<sup>−/−</sup> mice consistently took longer to find the hidden platform compared to their wild-type littermates (Bonferroni post-hoc test) (Fig. 2C). Two-way ANOVA revealed a significant effect of genotype (*F*<sub>1,98</sub> = 34.25, *P* < 0.001) and day of training (*F*<sub>7,98</sub> = 9.043, *P* < 0.001). When the platform was removed during the probe trial, *Bmal1*<sup>−/−</sup> littermates spent significantly more time in the quadrant where the platform was during training compared to *Bmal1*<sup>+/+</sup> mice, 45.4 ± 3.9% vs. 20.3 ± 5.0% of probe trial (*F*<sub>3,52</sub> = 4.080, *P* < 0.01, two-way ANOVA with Bonferroni post-hoc test) (Fig. 2D). Furthermore, *Bmal1*<sup>+/+</sup> mice crossed the platform’s previous location more frequently than *Bmal1*<sup>−/−</sup> mice, 3.9 ± 0.8 vs. 0.8 ± 0.4 crossings (*t*<sub>13</sub> = 2.964, *P* = 0.0110, two-tailed unpaired *t*-test).

While *Bmal1*<sup>−/−</sup> mice showed impairment in contextual fear conditioning and the Morris water maze, they demonstrated no deficit in the novel object recognition task. Mice were exposed to object A for 5 min, three times. Both *Bmal1*<sup>+/+</sup> and *Bmal1*<sup>−/−</sup> mice spent more time exploring novel object B 30 min after training (*F*<sub>5,32</sub> = 18.12, *P* < 0.001, one-way ANOVA, Tukey post-hoc test) (Fig. 2E). In fact, *Bmal1*<sup>−/−</sup> mice even spent more time interacting with a novel object 24 h after exploring the original object (*P* < 0.001, Tukey post-hoc test) (Fig. 2E).

*Bmal1*<sup>−/−</sup> mice have impaired LTP

The Schaffer collateral efferent fibers in the stratum radiatum of hippocampal area CA1 were stimulated in the acute brain slice and the resulting fEPSPs recorded. Before inducing LTP, the input/output (I/O) curve and paired-pulse facilitation were characterized. There was no significant difference between the *Bmal1*<sup>+/+</sup> and *Bmal1*<sup>−/−</sup> I/O values at each stimulus (Tukey’s multiple comparison test) (Fig. 3A). There was also no significant difference between the normalized slopes of the second fEPSP at each interpulse interval for paired-pulse facilitation (Tukey’s multiple comparison test) (Fig. 3B). To induce LTP, three tetanic stimuli separated by 5 min were delivered to the acute slice and fEPSPs monitored for 2 h (Fig. 3C). For the first 30 min following LTP induction, the *Bmal1*<sup>+/+</sup> hippocampus mean fEPSP slope response was 251.50 ± 3.43% of baseline, while the *Bmal1*<sup>−/−</sup> hippocampus demonstrated significantly reduced LTP with a mean fEPSP slope response of 152.20 ± 2.51% of baseline (*U* = 0.0, *P* < 0.0001, *r* = 1.21, Mann–Whitney). The *Bmal1*<sup>−/−</sup> hippocampus continued to demonstrate a significantly reduced response (228.7 ± 1.2% of baseline vs.

![Figure 1.](https://www.learnmem.org/figure1.png)
Figure 2. Bmal1−/− mice demonstrate impaired learning and memory in hippocampus-dependent tasks. (A) Contextual fear freezing behavior at ZT4. Bmal1−/− mice freeze significantly less 24 h (F(3,45) = 31.44, P < 0.0001) after contextual fear conditioning training with a 0.7-mA foot shock. One-way ANOVA with Tukey post-hoc test. (B) Contextual fear freezing behavior at ZT16. Both groups demonstrate significant freezing behavior 30 min and 24 h after training (F(3,41) = 17.49, P < 0.0001, one-way ANOVA, Tukey post-hoc test). (C) Morris water maze training curve. Bmal1−/− consistently took longer to find the platform for four trials of training per day over 8 d. There is a significant effect of genotype (F(1,96) = 34.25, P < 0.001) and day of training (F(2,96) = 9.043, P < 0.001). Two-way ANOVA with Bonferroni post-hoc test. (D) Morris water maze probe trial. Bmal1+/+ mice spend significantly more time in quadrant 3, where the platform used to be, compared to other quadrants and Bmal1−/− mice (F(3,52) = 4.080, P < 0.01). Two-way ANOVA with Bonferroni post-hoc test. (E) Novel object recognition. Thirty minutes after training to object A, both Bmal1−/− and Bmal1+/+ mice spend significantly more time exploring novel object B (F(3,22) = 18.12, P < 0.001). Bmal1−/− mice spent more time interacting with novel object C 24 h after exposure to object A (P < 0.001). One-way ANOVA, Tukey post-hoc test. All values represent mean ± SEM, n = 6–10.

133.3 ± 1.1% of baseline) 60–90 min following induction (U = 0.0, P < 0.0001, r = 1.22, Mann–Whitney). These results are consistent with behavioral data, suggesting a reduction in long-term synaptic plasticity.

The hippocampus of Bmal1−/− mice does not demonstrate diurnal oscillation of MAPK activity or cAMP

Previous studies have shown that wild-type mice exhibit a diurnal oscillation in MAPK activity and cAMP that is required for long-term hippocampus-dependent memory (Eckel-Mahan et al. 2008). Therefore, Bmal1+/+ and Bmal1−/− hippocampi were collected at ZT4 and ZT16, the peak and trough of the MAPK oscillation, and analyzed for MAPK activity. Two-way ANOVA revealed significant effects of genotype (F(1,26) = 6.085, P < 0.05) and time of day (F(1,26) = 6.875) on diurnal phospho-MAPK levels. Bmal1+/+ hippocampi demonstrated increased levels of phospho-MAPK at ZT4 compared to ZT16 (P < 0.05, Bonferroni post-hoc test) (Fig. 4B). Bmal1−/− hippocampi showed no diurnal change in phospho-MAPK at ZT4 and exhibited lower levels of phospho-MAPK at ZT4 compared to Bmal1+/+ hippocampi (P < 0.05, Bonferroni post-hoc test) (Fig. 4B). Similarly, two-way ANOVA revealed a significant effect of genotype (F(1,54) = 21.56, P < 0.0001) and time of day (F(2,54) = 9.101, P = 0.0039) on diurnal cAMP levels. Bmal1−/− hippocampi demonstrated increased levels of cAMP at ZT4 compared to ZT16 (P < 0.05, Bonferroni post-hoc test) (Fig. 4C). Bmal1−/− hippocampi showed no diurnal change in cAMP at ZT4 and exhibited lower levels of cAMP at ZT4 compared to Bmal1+/+ hippocampi (P < 0.001, Bonferroni post-hoc test) (Fig. 4C).

The Bmal1−/− hippocampus shows no training-induced activation of MAPK

When wild-type mice are trained for contextual fear memory, there is a measurable increase in MAPK activity in area CA1 of the hippocampus (Sindreu et al. 2007). Therefore, Bmal1+/+ and Bmal1−/− mice were exposed to a novel context (ZT4 to ZT16) and either immediately received a 0.7-mA foot shock or were first allowed to explore for 2 min. Bmal1+/+ mice that experienced the paired association between the novel context and foot shock showed an increase in phospho-MAPK in the hippocampus 30 min after training compared to unpaired controls which were shocked immediately (Bmal1−/− hippocampi t(16) = 2, P = 0.0121, two-tailed unpaired t-test) (Fig. 5). There was no difference between Bmal1+/+ paired and unpaired hippocampi (t(15) = 1.585, P = 0.1339, two-tailed unpaired t-test) (Fig. 5), indicating that MAPK activity did not increase when Bmal1−/− mice were trained for contextual fear memory.

Discussion

Bmal1−/− mice offer the opportunity to investigate the relationship between a functional molecular clock and memory. We discovered that Bmal1−/− mice are impaired in hippocampus-dependent memory tasks. At 24 h after training for contextual fear conditioning, Bmal1−/− mice freeze less than wild-type littermates when trained and tested during the day. While Bmal1+/− mice do learn and exhibit fear memory for the foot shock, their memory for contextual fear is similar to that of wild-type littermates during the night. Bmal1−/− mice are also impaired in spatial learning and memory, as measured by the Morris water maze. Consistent with these memory deficits, LTP at the Schaffer-collateral pathway is reduced in the Bmal1−/− hippocampus. The Bmal1−/− hippocampus also does not demonstrate
Figure 3. LTP at the Schaffer-collateral synapse is impaired in the Bmal1\textsuperscript{-/-} hippocampus. (A) Input/output (I/O) relationship between stimulus (µA) and the slope of the postsynaptic evoked fEPSP (mV/ms). Stimulus ranged from 20 to 150 µA. There is no difference between the Bmal1\textsuperscript{+/+} and Bmal1\textsuperscript{-/-} I/O values at each stimulus. Tukey’s multiple comparison test. n = 10–12. (B) Paired-pulse facilitation with interpulse interval ranging from 20 to 290 msec. There was no difference between the normalized slopes of the second iEPSP at each interpulse interval. Tukey’s multiple comparison test. n = 7–9. (C) Averaged LTP response of the Schaffer-collateral fibers, represented by fEPSP slope and normalized to pretetanic baseline. To induce LTP, three tetanic stimuli (1-sec, 100-Hz train with 0.1-msec pulse length) separated by 5 min were delivered. Bmal1\textsuperscript{-/-} hippocampus demonstrated significantly reduced LTP with a mean fEPSP slope response within first 30 min following induction (U = 0.0, P < 0.0001, r = 1.21, Mann–Whitney). The Bmal1\textsuperscript{-/-} hippocampus continued to demonstrate a significantly reduced response 60–90 min following induction (U = 0.0, P < 0.0001, r = 1.22, Mann–Whitney). (D) Representative LTP traces from individual slices. Black is 5 min after tetanic stimulations. Red is 60 min after tetanic stimulations. All values represent mean ± SEM.
There is no diurnal oscillation of ERK activity or cAMP in Bmal1+/− hippocampi. (A) Western blot image depicts representative phosphorylated ERK and ERK intensities from isolated hippocampi of individual mice at ZT4 or ZT16. (B) Ratio of pERK to ERK volume intensities normalized to control Bmal1+/−/ZT4 levels. There is a significant effect of genotype ($F_{1,26} = 6.085$, $p < 0.05$) and time of day ($F_{1,26} = 6.875$) on diurnal phospho-MAPK levels. For Bmal1+/− hippocampi, phospho-MAPK is higher at ZT4 compared to ZT16 ($p < 0.05$). Bmal1+/− hippocampi showed no change in phospho-MAPK activity between ZT4 and ZT16 and exhibited lower levels of phospho-MAPK at ZT4 compared to Bmal1−/− hippocampi ($p < 0.05$). Two-way ANOVA with Bonferroni post-hoc test. All values represent mean ± SEM, $n = 6–9$. (C) Ratio of cAMP concentration (pmol) to protein (mg) normalized to Bmal1+/−/ZT4 group. There are significant effects of genotype ($F_{1,54} = 21.56$, $p < 0.0001$) and time of day ($F_{1,54} = 9.101$, $p = 0.0039$) on diurnal cAMP levels. Bmal1+/− hippocampi demonstrated increased levels of cAMP at ZT4 compared to ZT16 ($p < 0.05$). Bmal1−/− hippocampi showed no diurnal change in cAMP at ZT4 and exhibited lower levels of cAMP at ZT4 compared to Bmal1+/− hippocampi ($p < 0.001$). Two-way ANOVAs with Bonferroni post-hoc test. All values represent mean ± SEM, $n = 12–16$.

We find that genetically abolishing the molecular clock compromises hippocampus-dependent learning and memory tasks. This report extends our understanding of the relationship between diurnal rhythms and the hippocampus and is consistent with substantial literature implicating the circadian system in memory (Gerstner and Yin 2010).

Materials and Methods

Mice

Bmal1+/− breeders in the C57/B6 background were generously provided by Dr. Christopher Bradfield from the University of Wisconsin Medical School, Madison, WI. Mice were genotyped with PCR as previously described (Bunger et al. 2000). Bmal1−/− littermates were used as controls. All experiments were performed on adult, male mice aged 3–5 mo. Food and water were provided ab libitum and animals were maintained on a 12-h light-dark (LD) cycle, with light onset ZT0 at 8 a.m., unless otherwise specified. Experiments were in accordance with the Institutional Animal Care and Use Committee’s recommendations at the University of Washington.

Activity monitoring

Mice were individually housed and their voluntary activity data were acquired using QA-4 activity input modules coupled to infrared motion detectors and the VitalView Data Acquisition System (Mini Mitte, version 4.1). Actograms and periodograms were generated with the ImageJ plug-in ActogramJ (University of Wisconsin). Mice were monitored for 7 d in 12-h LD conditions and 14 d in DD conditions. Only the last 7 d in DD conditions were included in depicted actograms and periodogram analysis.

Contextual fear conditioning

Mice were handled for at least 5 d before all behavioral testing. For contextual fear training, mice were exposed to a 10×10×10 cm (L×W×H) transparent Plexiglas chamber with a metal shock grid (Colburn Instruments). Between each subject, the chamber was wiped with diluted acetic acid. Mice were allowed to explore the novel context for 2 min before receiving a 0.7-mA foot shock. They were returned to their home cage after a 1-min recovery. Behavior during testing was video recorded and the percentage of freezing activity was scored by a blinded investigator. Freezing was scored as all four paws on the ground and no movement other than respiration. Mice were trained and tested from ZT3 to ZTS, during the light phase, or under red light from ZT15 to ZT17, during the night phase.

Morris water maze

Mice were placed in a 122-cm-diameter pool filled with room temperature water (22°C) made opaque with nontoxic, white, tempera paint. They were trained for 8 d, four times a day, to locate the hidden and fixed platform (circular, 5-cm diameter) using visual cues to navigate. Mice were placed in randomized locations of the pool for each trial. During training, the mice were given 90 sec to find the platform and guided to the platform when they did not find it. They sat on the platform for 30 sec before removal. Their memory was tested using a probe trial, where the platform is removed, lasting 90 sec. The probe trial was video recorded and their swim behavior was analyzed using Ethovision (Noldus). Mice were trained and tested from ZT3 to ZT8, during the light phase.

Novel object recognition

Mice were habituated to a 17 (W) × 8.5 (D) × 6 (H) cage overnight. After habituation, they explored two identical plastic objects (A) for 5 min, three times, in the habituated cage. Thirty minutes after training, they were presented with object A and a new object B for 5 min. Twenty-four hours after training, they were presented with object A and a new object C for 5 min. Objects were sprayed with ethanol in between animal subjects. Investigators video recorded each trial and later scored exploration time with each object. Exploration was defined as anytime a mouse’s head was oriented toward the object and their nose was within 1 cm of the object. Time spent climbing on objects was eliminated from exploration time. Mice were trained and tested from ZT3 to ZT16, during the light phase.

Electrophysiology

Hippocampal LTP was compared between Bmal1+/− and Bmal1−/− male mice at ages 2–3 mo. After dissection at ZT2, brains were put in ice-cold cutting aCSF (12.5 mM NaCl, 0.25 mM KCl, 0.6 mM MgCl2, 0.05 mM CaCl2, 0.125 mM NaH2PO4, 10 mM glucose, 25 mM NaHCO3) containing APV (40 μM, obtained from Tocris) and then cut into 400-μm coronal sections using a vibratome (Leica VT1000S, Leica Microsystems). Slices recovered for 2 h in aCSF (12.5 mM NaCl, 0.25 mM KCl, 0.1 mM MgCl2, 0.25 mM CaCl2, 0.125 mM NaH2PO4, 10 mM glucose, 25 mM NaHCO3). Slices were continuously

Figure 4. There is no diurnal oscillation of ERK activity or cAMP in Bmal1−/− hippocampi. (A) Western blot image depicts representative phosphorylated ERK and ERK intensities from isolated hippocampi of individual mice at ZT4 or ZT16. (B) Ratio of pERK to ERK volume intensities normalized to control Bmal1+/−/ZT4 levels. There is a significant effect of genotype ($F_{1,26} = 6.085$, $p < 0.05$) and time of day ($F_{1,26} = 6.875$) on diurnal phospho-MAPK levels. For Bmal1+/− hippocampi, phospho-MAPK is higher at ZT4 compared to ZT16 ($p < 0.05$). Bmal1−/− hippocampi showed no change in phospho-MAPK activity between ZT4 and ZT16 and exhibited lower levels of phospho-MAPK at ZT4 compared to Bmal1−/− hippocampi ($p < 0.05$). Two-way ANOVA with Bonferroni post-hoc test. All values represent mean ± SEM, $n = 6–9$. (C) Ratio of cAMP concentration (pmol) to protein (mg) normalized to Bmal1+/−/ZT4 group. There are significant effects of genotype ($F_{1,54} = 21.56$, $p < 0.0001$) and time of day ($F_{1,54} = 9.101$, $p = 0.0039$) on diurnal cAMP levels. Bmal1+/− hippocampi demonstrated increased levels of cAMP at ZT4 compared to ZT16 ($p < 0.05$). Bmal1−/− hippocampi showed no diurnal change in cAMP at ZT4 and exhibited lower levels of cAMP at ZT4 compared to Bmal1+/− hippocampi ($p < 0.001$). Two-way ANOVAs with Bonferroni post-hoc test. All values represent mean ± SEM, $n = 12–16$.
oxygenated with a biological mix (95% oxygen, 5% carbon dioxide). After recovery, the Schaffer collateral efferent fibers in the stratum radiatum of hippocampal area CA1 were stimulated and the resulting fEPSPs recorded from ZT4 to ZT10. The slice was stimulated with the S88 Stimulator (Astro-Med Inc., Grass Instrument Division) using a concentric bipolar electrode (Frederick Haer & Co). Electric responses were amplified by the Axopatch 200B (Axon Instruments) and digitized by the Digidata 1440A (Axon Instruments). Before inducing LTP, the basal field fEPSP activity was developed with the CDP-star western alkaline phosphatase-conjugated goat to mouse IgG (Sigma). Immunoreactivity was evaluated with the CDP-star western alkaline phosphatase detection system (Tropix). Blots were imaged with ChemiDoc XR+ and analyzed with Image Lab (BioRad). pERK intensity was normalized to ERK intensity, which served as a loading control.

cAMP ELISA

The ELISA-based cAMP Biotek Enzyme immunoassay System protocol (Amersham Bioscences) was used with some minor deviations. Hippocampal tissue was collected and homogenized as described in the Western methods but with the addition of 1 mM of the PDE inhibitor IBMX. Ice-cold ethanol (100%) was added to the homogenate. Homogenates were spun for 2 min at a speed of 1000g at 4°C. Resulting supernatants were evaporated in a heat block at 55°C and precipitates were resuspended in 100 μL assay buffer. Competition binding was carried out according to the Enzyme Immunoassay System instructions. Extracts were diluted 1:30 to achieve concentrations in the linear range of the assay. cAMP levels were normalized to protein concentrations.

Statistical analysis

Statistical tests were performed in GraphPad Prism. α value was set at 0.05. The specific tests used are reported with results.

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References

show normal acquisition but impaired long-term retention of place information in the water task. *Neurobiol Learn Mem* **75**: 51–62.


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In the aforementioned article, the second author’s name has been corrected to Trongha X. Phan, as noted above. The PDF and full-text versions online have been updated to reflect this change as well.
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